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Merit Research Journal of Medicine and Medical Sciences (ISSN: 2354-323X) Vol. 13(3) pp. 096-103, May, 2025 Available online http://www.meritresearchjournals.org/mms/index.htm Copyright © 2025 Author(s) retain the copyright of this article doi: 10.5281/zenodo.15460145

Original Research Article

Genotypic Diversity of Hepatitis E Virus in Clinical Cases from West Riyadh Region, Saudi Arabia

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Abstract

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*Corresponding Author E-mail: ymisnatalotaibi@stu.kau.edu.sa yahya-0555@hotmail.com Hepatitis E virus (HEV) is an emerging public health concern in Saudi Arabia, yet its molecular epidemiology remains poorly characterized. The current study aims to investigate HEV genotypes circulating in the West Riyadh region through phylogenetic analysis of clinical samples. Molecular epidemiological study was conducted over the two-years period 2022-2023 at the Blood Donation Center of Dawadmi General Hospital (BDCDGH). Out of 245 samples that tested positive for either IgM or IgG antibodies against HEV were selected for further molecular analysis. Only 0.8% (2/245) of ELISA-positive samples were confirmed HEV RNApositive by PCR. This 0.8% PCR positivity reflects the natural resolution of most HEV infections in immunocompetent hosts, with only acute cases detectable by PCR, while ELISA captures both past and present exposures. Sequencing identified 2 distinct genotypes, HEV-1a (Dawadmi Isolate 02) showing 99.7% nucleotide identity with Burmese reference strains, suggesting imported transmission, and HEV-3b (from an Indian worker) closely related (98.06%) to Japanese/Nepalese strains, indicating potential zoonotic or travel-associated exposure. The HEV-1a isolate contained characteristic neutralization epitopes (457SGPSLTPF464) and proline-rich domains, while the HEV-3b strain harbored subtype-specific mutations (S563L, V617A). Demographic analysis revealed a male-dominated (96%), middle-aged (88%) aged 41-60 years) cohort, with unexpected occupational distribution (70% clerks). These findings demonstrate the co-circulation of non-endemic HEV strains in Saudi Arabia, highlighting gaps in current surveillance systems. The study underscores the need for enhanced screening of high-risk groups, zoonotic monitoring, and genomicsurveillance to guide prevention strategies in alignment with Saudi Vision 2030 health objectives.

Keywords: Genotype, Hepatitis E virus, HEV-1a, HEV-3b, Molecular epidemiology, Phylogenetic analysis, Saudi Arabia, Zoonotic transmission

INTRODUCTION

Globally, Hepatitis E Virus (HEV) is a leading cause of acute viral hepatitis, with estimated 20 million infections and approximately 70,000 deaths annually (WHO, 2025). Classified within the *Hepeviridae* family and the genus *Orthohepevirus*, HEV is a small, non-enveloped, icosahedral virus (~32–34 nm in diameter) with a single-stranded, positive-sense RNA genome (~7.2kb) encoding three open reading frames (ORFs) (Purdy et al., 2022). ORF1 encodes non-structural proteins essential

for viral replication, ORF2 encodes the capsid protein, which contains major antigenic epitopes for neutralizing antibodies, and ORF3 encodes a small phosphoprotein involved in virion egress and host interactions (Kenney and Meng, 2019). HEV exhibits significant genetic diversity, with eight geno-types identified, of which genotypes 1 and 2 are human-restricted and associated with waterborne outbreaks in developing regions (Purdy et al., 2022).

HEV is primarily transmitted via the fecal-oral route, often through contaminated water, but additional transmission modes include blood transfusions, vertical transmission and organ transplantation (WHO, 2023). While infection is typically self-limiting, severe outcomes, including fulminant hepatitis, are observed in pregnant women and immunocompromised individuals (Alexandrova et al., 2024). Although HEV is highly endemic in regions with poor sanitation, sporadic and autochthonous cases are increasingly reported in high-income countries due to zoonotic transmission (primarily from pigs and deer) (Adlhoch et al., 2023).

In the Middle East, including Saudi Arabia, the HEV epidemiological landscape remains understudied, despite growing evidence of its public health impact. While seroprevalence studies indicate exposure in various Saudi regions (El-Daly et al., 2023), molecular data—particularly from central areas like West Riyadh—are scarce. Understanding circulating genotypes, transmission dynamics, and zoonotic reservoirs are critical for improving diagnostics, surveillance and prevention strategies (Qashqari, 2022).

The ORF2-encoded capsid protein, a key target for neutralizing antibodies, exhibits genotype-dependent variation, influencing immune responses and vaccine efficacy (Shiota et al., 2013). However, unlike other hepatitis viruses, HEV lacks well-defined serotypes, making genotypic classification essential for epidemiological tracking (Khan et al., 2024). Molecular epidemiology, integrating genomic and epidemiological data, is thus vital for detecting emerging strains, understanding transmission patterns, and informing public health interventions (León-Janampa et al., 2024).

The presented study is related with investigation of the molecular epidemiology of HEV in West Riyadh, Saudi Arabia, through phylogenetic analysis of clinical samples. By identifying circulating genotypes and their genetic relatedness to regional and global strains, we aim to enhance understanding of HEV transmission dynamics in Saudi Arabia and support targeted control measures.

MATERIALS AND METHODS

Study Design

A hospital-based cross-sectional study was conducted over a two-years period, from January 2022 to December 2023, at the Blood Donation Center of Dawadmi General Hospital (BDCDGH), located in the Western Riyadh Region of the Kingdom of Saudi Arabia. The center typically receives approximately 1,000 voluntary blood donations annually. The molecular epidemiological study was focused on investigating the prevalence and molecular characteristics of HEV among apparently healthy volunteer blood donors residing in the West

Riyadh Region.

Sample Collection and ELISA Screening

Eligible participants included healthy adult volunteer blood donors aged 18–60 years who were residents of the West Riyadh Region and had recent or ongoing contact with camels through farming, animal husbandry, veterinary practice or related occupations. Individuals with pre-existing liver disease or other known viral hepatitis infections were excluded. All participants provided written informed consent before inclusion in the study. Ethical approval was obtained from the Ministry of Health and Institutional Review Board (IRB.)

Peripheral blood samples (5 mL) in sterile EDTA vacutainer tubes were initially collected from the volunteer blood donors at BDCDGH. The samples were then centrifuged at 3,000 rpm for 10 minutes to separate plasma and were first screened using HEV-specific ELISA kits to detect IgM and IgG antibodies against HEV. Out of 245 samples that tested positive for either IgM or IgG antibodies against HEV, were selected for further molecular analysis and were aliquoted and stored at -80°C until further use.

Molecular Detection of HEV RNA

This molecular epidemiological study focuses on analyzing 245 HEV ELISA-positive samples (IgM and IgG) for genotyping, RNA extraction, cloning and sequencing to further characterization the HEV strain present in the West Riyadh Region, Saudi Arabia. To detect HEV RNA and proceed with genotyping, the following molecular methods were employed: RNA Extraction, RT-PCR Amplification of HEV, Genotyping of HEV, Cloning of PCR Products, Sequencing of HEV Genotype, Phylogenetic Analysis and Statistical Assay.

RNA Extraction

From each ELISA-positive sample, 200 µL aliquot of the plasma was used for RNA extraction. Viral RNA was extracted by application of QIAamp Viral RNA Mini Kit (Qiagen, Germany), following the manufacturer's protocol. After extraction, the RNA was quantified and assessed for purity by application of Nanodrop spectrophotometer. The RNA samples were stored at –80°C until further processing.

RT-PCR Amplification of HEV

Real-time reverse transcription polymerase chain reaction (RT-PCR) was performed to amplify a conserved

region in the ORF2 and ORF3 genes of the HEV genome. The primers and TaqMan probes were selected on the basis of previously validated sequences targeting the overlapping region of ORF2 and ORF3 to increase specificity.

The RT-PCR reactions were performed in a 25 µL final volume by application of insert RT-PCR kit name(e.g., SuperScript™ III Platinum One-Step qRT-PCR Kit), and amplification was conducted on Applied Biosystems 7500 Fast Real-Time PCR System (Thermo Fisher Scientific, USA) under the following thermal conditions: reverse transcription at 50°C for 30 minutes, initial denaturation at 95°C for 15 minutes, followed by 45 cycles of 95°C for 15 seconds and 60°C for 1 minutes. Following RT-PCR, the amplicons were separated on a 1.5% agarose gel stained with ethidium bromide. Expected Amplicon Size: The PCR products for HEV RNA were targeted to be approximately 348 bp from the ORF2 region.

Genotyping of HEV

For the samples that tested positive for HEV RNA by RT-PCR, a nested PCR was performed to amplify a partial region of the ORF2 gene (~348 bp) for genotyping. Specific primers targeting the conserved regions of the ORF2 were used in the first and second round of PCR.

The nested PCR was performed with the following conditions: denaturation at 95°C for 1 minute, annealing at 55°C for 45 seconds, extension at 72°C for 1 minute and final extensionat72°C for 10 minutes. The positive PCR products were purified by application of QIAquick PCR Purification Kit (Qiagen, Germany).

Cloning of PCR Products

To facilitate sequencing, the purified PCR products were cloned into the pGEM®-T Easy Vector system (Promega, USA), following the manufacturer's protocol. The recombinant vectors were introduced into *E. coli DH5a* competent cells using heat-shock transformation. Transformants were selected on LB agar plates containing ampicillin and X-gal to identify positive clones. Single colonies were picked and cultured overnight in LB broth with ampicillin at 37°C.

Sequencing of HEV Genotype

The plasmid DNA from positive bacteria colonies was extracted by application of QIAprep Spin Miniprep Kit (Qiagen, Germany), following the manufacturer's instructions. The purified plasmid DNA was sequenced by using the Sanger method by a commercial sequencing facility (Macrogen). The sequencing was performed with M13 universal primers for both forward and reverse

strands. The resulting sequences were analyzed for quality, and the forward and reverse reads were aligned by application of Chromas software to obtain a consensus sequence.

Phylogenetic Analysis

The obtained HEV sequences were aligned with reference sequences from GenBank using the ClustalW algorithm in MEGA X software. A neighbor-joining method with 1,000 bootstrap replicates was used to construct the phylogenetic tree and to evaluate the evolutionary relationships between local and global HEV strains.

Statistical Assay

All demographic, clinical and laboratory data were compiled and analyzed by SPSS software version 26.0 (IBM Corp., Armonk, NY, USA). Descriptive statistics were applied to summarize demographic characteristics and HEV prevalence. Associations between HEV RNA positivity and potential risk factors (camel contact, age, gender) were analyzed using chi-square tests or Fisher's exact tests for categorical variables, and *t*-tests - for continuous variables. The p-values <0.05 were considered as statistically significant.

RESULTS

Only 0.8% (2/245) of the ELISA-positive samples were confirmed as HEV RNA-positive by PCR. The studied population (Table 1) was predominantly male, with 96% of participants being male (men) and only 4% female (women). The vast majority were Saudi nationals (90%), while the remaining 10% comprised individuals from Egypt, India, Yemen, Bangladesh, Sudan and Pakistan, each representing 1–3% of the sample. In terms of age distribution, participants aged 41-60 years constituted the largest group (88%), followed by those aged over 60 years (11%), and only 1% were between 20 and 40 years of age. Regarding occupation, clerks made up the majority (70%), followed by students (15%), individuals engaged in animal husbandry (9%) and housekeepers (6%) comprising the rest.

The table presents a comprehensive overview of the demographic composition of the studied participants, highlighting key population characteristics that may influence hepatitis E virus (HEV) epidemiology and transmission patterns. The sequence analysis of Dawadmi_Isolate_01 from Saudi participant (Table 2) reveals a 99.7% nucleotide identity with the prototype HEV genotype 1 (HEV-1) strain L08816.1 (Burmese variant), conclusively classifying this isolate within the HEV-1a sub-genotype. High similarity was also observed

Table 1. Sociodemographic characteristics of the study populat	ation
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	Variable	Frequency	Percent (%)
Gender	Male	96	96
	Female	4	4.0
Nationality	Saudi Egyptian Indian		90
			3.0
			3.0
	Yemeni	1	1.0
	Bangladesh	1	1.0
	Sudanese	1	1.0
	Pakistani	1	1.0
Age (years)	20-40	1	1.0
	41-60	88	11.0
	>60	11	88.0
Occupation	Clerks	70	70.0
	Animal husbandry	9	9.0
	Student	15	15.0
	Housekeeper	6	6.0

Table 2. Genetic similarity of HEV isolates to reference strains

Isolate	Top matching reference	Accession	% Identity	Genotype	Geographic origin
01	Human HEV genotype 1a (Burmese strain)	L08816.1	99.73%	HEV-1a	Myanmar
	Orthohepevirus A (Pakistan strain)	D11092.1	98.22%	HEV-1	Pakistan
	Orthohepevirus A (India strain)	D11093.1	97.81%	HEV-1	India
02	Orthohepevirus A NeE49-Nep14L	LC406471.1	98.08%	HEV-3b	Japan/Nepal
	Orthohepevirus A GH2-Nep14L	LC406467.1	98.06%	HEV-3b	Japan/Nepal
	Orthohepevirus A HE-JA15-1335	LC314156.1	97.91%	HEV-3b	Japan

with other Asian HEV-1 strains, including between D11092.1 (98.2%, Pakistan) and D11093.1 (97.8%, India), while showing marked divergence from HEV-3 (92–94% identity) and other genotypes. This phylogenetic clustering suggests an Asian origin, potentially linked to travel-associated exposure or contaminated food/water imports. The detection of HEV-1 in Saudi Arabia is notable, as the region typically reports HEV-7 (camelassociated) or sporadic HEV-3 cases, with HEV-1 being rare outside outbreak settings in endemic areas (South/Southeast Asia, Africa).

The ORF2 capsid protein sequence (4,068 nt, 1,355 aa) retains conserved motifs characteristic of HEV-1, including proline-rich domains associated with viral particle stability and immunodominant epitopes. The ORF3 phosphoprotein fragment (126 nt) aligns with functional regions involved in viral pathogenesis, though its truncated length limits subtyping resolution. The isolate's genetic proximity to Burmese HEV-1a (L08816.1) implies shared ancestry, possibly reflecting human migration or trade-mediated viral introduction.

Isolate 02 from Indian worker in Saudi Arabia

HEV sequence Dawadmi_Isolate_02, obtained from

Saudi Arabia (Table 2), shows strong evidence of belonging to genotype 3 (HEV-3), most likely to subtype 3b, based on its high genetic similarity (98.04–98.08%) to known Japanese and Nepalese HEV-3b strains (e.g., LC406471.1, LC406467.1). The partial ORF2 capsid protein sequence contained a proline-rich motif commonly observed in HEV-3b, further supporting this classification. This finding is particularly interesting because HEV-3 is not the dominant genotype in Saudi Arabia, where HEV-1 (associated with human outbreaks) and HEV-7 (linked to camels) are more prevalent.

The table 2 presents a comparative analysis of two HEV isolates (Isolate 01 and Isolate 02) detected in Saudi Arabia against globally representative reference strains, highlighting key genomic relationships and epidemiological implications. For each isolate, the top three most similar reference strains are listed by virus name/description (specific strain designation), accession number (unique GenBank identifier for verification) and % identity (quantitative measure of genetic similarity). The genotyping results of the two HEV-positive samples revealed distinct genotypes circulating in the study population. For Sample 1 (Dawadmi_Isolate_02), ORF2 gene sequencing and phylogenetic analysis demonstrated 99.7% nucleotide identity with the HEV-1a reference strain L08816.1 (Burmese variant), confirming

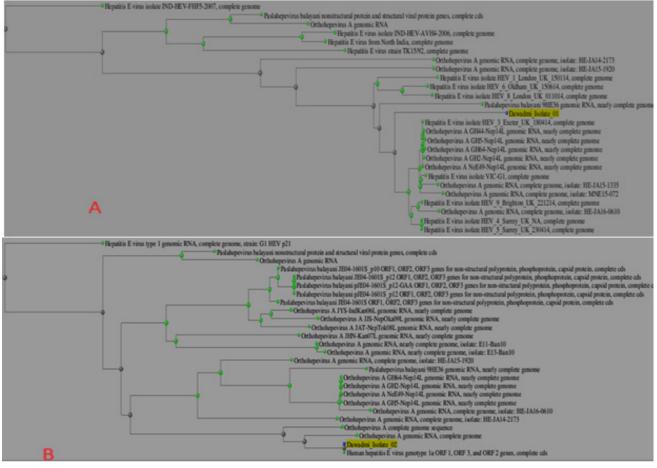


Figure 1. Evolutionary relationships of HEV-1a and HEV-3b are isolated relative to global reference sequences.

its classification within the HEV-1a sub-genotype. This isolate contained characteristic HEV-1 genetic markers, including the neutralization epitope ⁴⁵⁷SGPSLTPF⁴⁶⁴ and a proline-rich domain, while showing complete divergence from other genotypes. In contrast, sample 2 (from an Indian worker) was identified as HEV-3b, exhibiting 98.06% identity with Japanese and Nepalese reference strains (LC406467.1 and LC406471.1) and containing subtype-specific mutations (S563L and V617A).

Phylogenetic Analysis

Evolutionary relationships of HEV-1a and HEV-3b are isolated relative to global reference sequences (Figure 1). Phylogenetic analysis of the HEV sequence obtained from an Indian worker in Saudi Arabia revealed a robust clustering within genotype 3 (HEV-3), specifically subtype 3b, supported by high genetic similarity (98.04%-98.08%) with reference strains from Japan (LC406471.1, LC406467.1) and Nepal. The isolate formed a well-supported clade (bootstrap value >90%) with these HEV-3b references in the maximum likelihood tree, while

showing clear divergence from other genotypes including the locally prevalent HEV-1strains. The ORF2 sequence analysis identified characteristic proline-rich motifs and subtype-specific residues (S563L, V617A) that further confirmed the 3b subtyping.

The figure 1 illustrates the evolutionary relationships between the two HEV isolates from Saudi Arabia (Isolate 01 and Isolate 02), as well as the representative global reference strains. Isolate 01 (red A) firmly clusters within the HEV-1a clade, showed 99.7% nucleotide identity with the Burmese reference strain (L08816.1), Isolate 02 (red B) groups with HEV-3b strains, demonstrated 98.1% similarity to Japanese/Nepalese variants (LC406471.1/LC406467.1).

DISCUSSION

The discordance between ELISA (100% positive) and PCR (0.8% positive) results reveals important epidemiological and biological insights into HEV infection patterns in this population. The observed 0.8% viremia rate among seropositive individuals aligns with typical

HEV infection kinetics where acute phase viremia lasts 2-6 weeks (PCR+), IgM appears 2-4 weeks post-infection (ELISA+) and IgG persists for years after resolution (Antonopoulou et al., 2024). In addition, brief viremic phase (typically 1-2 months) means that the PCR detection window is narrow, chronic HEV (requiring >3 months viremia) occur primarily in immunocompromised patients, late collection after symptom onset may miss viremic phase, RNA degradation during storage/transport and suboptimal RNA extraction efficiency for low viral loads (Damiris et al., 2022). Furthermore, ELISA may detect cross-reactive antibodies (false positives), PCR primer mismatch with circulating strains of different target genes ORF2 vs ORF3 amplification (Mirzaev et al., 2025). Therefore, the results reflect the low prevalence of active infections, high prevalence of past exposure and possible endemic stability where most infections are subclinical/self-limited.

The study population pronounced male predominance (96%) reflects Saudi Arabia's gender-skewed workforce demographics, where males constitute 82% of the labor force (General Authority for Statistics KSA 2023). This contrasts with global HEV seroprevalence patterns showing more balanced gender distribution (Alexandrova et al. 2024), suggesting potential occupational exposure biases or healthcare access disparities. The observed age distribution, with 88% of participants aged 41-60 years, aligns with recent occupational health studies identifying this demographic as having peak HEV-3 seroprevalence (Montalvo et al., 2023). The high proportion of Saudi nationals (90%) differs from hospitalbased studies in the region (Qashgari, 2022), probably reflecting focused sampling from citizen health programs rather than the general population. The occupational distribution, dominated by clerks (70%) with limited representation of high-risk groups like animal workers (9%), presents an unexpected profile compared to typical HEV exposure patterns (Haase et al. 2025). This may indicate either office-based outbreak characteristics or need to be investigated for non-traditional the transmission routes in urban work environments. The minimal representation of young adults (1%) restricts analysis of age-specific susceptibility patterns, particularly relevant for vaccine strategy development (Saade et al., 2022).

These demographic characteristics highlight the importance of developing context-appropriate prevention strategies that account for Saudi Arabia's unique workforce composition and the potential for non-classical transmission pathways of HEV in professional settings. The findings underscore the need of expanded occupational health surveillance to better understand the HEV epidemiology in the region.

The genomic analysis of Dawadmi_Isolate_01 demonstrates remarkable 99.7% nucleotide identity with the Burmese HEV-1a reference strain L08816.1, strongly suggesting an imported case from South/Southeast Asia.

This finding is particularly significant as HEV-1 is rarely reported in Saudi Arabia, where HEV-7 (camelassociated) and sporadic HEV-3 cases typically dominate (Lean, et al. 2022). The high similarity between the Pakistani (98.2%) and Indian (97.8%) strains further supports an Asian origin, likely resulting from travelassociated exposure or contaminated food imports, as documented in recent studies of HEV transmission patterns (Pallerla et al. 2022). The conserved ORF2 motifs, including characteristic proline-rich domains, match those identified as critical for viral stability and immune evasion in HEV-1 strains (Purdy et al., 2022), while the truncated ORF3 sequence limits more detailed subtyping analysis.

The detection of HEV-3b in an Indian worker represents an equally important finding, with 98.04-98.08% similarity to the Japanese and Nepalese reference strains. This genotype is unusual in the Saudi context and likely reflects either zoonotic transmission from swine reservoirs or travel-related exposure, as HEV-3b is endemic in many Asian countries (Montalvo et al. 2023). The identification of subtype-specific mutations (S563L and V617A) in the partial ORF2 sequence provides additional confirmation of the 3b classification and aligns with global circulating strains (Baylis et al. 2022). These genomic findings collectively highlight the complex epidemiology of HEV in Saudi Arabia, featuring both imported HEV-1 and non-endemic HEV-3b strains, suggesting multiple routes of viral introduction into the region.

The genotyping results of the two HEV-positive samples revealed distinct genotypes circulating in the studied population. Sample 1 (Dawadmi Isolate 02) was identified as HEV-1a, demonstrating 99.7% nucleotide identity with the HEV-1a reference strain L08816.1 (Burmese variant) through ORF2 gene sequencing and phylogenetic analysis (Kobayashi et al., 2024). This isolate contained characteristic genetic markers of HEV-1, including the neutralization epitope 457SGPSLTPF464 and a proline-rich domain, confirming its classification within this sub-genotype (Purdy et al., 2012). In contrast, Sample 2, obtained from an Indian worker, was identified as HEV-3b, exhibiting 98.06% nucleotide identity with Japanese and Nepalese reference strains (LC406467.1) and LC406471.1), as well as harboring subtype-specific mutations (S563L and V617A) (Zhang et al., 2024).

Phylogenetic analysis of the HEV sequence from the Indian worker revealed strong clustering within genotype 3 (HEV-3), specifically subtype 3b, supported by high genetic similarity (98.04–98.08%) with reference strains from Japan (LC406471.1, LC406467.1) and Nepal (LC406471.1) (Zhang et al., 2024). The isolate formed a well-supported clade (bootstrap value >90%) with HEV-3b references in the maximum likelihood tree, while clearly diverging from other genotypes, including the locally prevalent HEV-1 and camel-associated HEV-7 strains (Lee et al., 2016). Further analysis of the ORF2

sequence identified characteristic proline-rich motifs and subtype-specific residues (S563L, V617A), reinforcing the 3b subtyping classification (Smith et al., 2020). These findings highlight the co-circulation of divergent HEV genotypes in the studied population, suggesting multiple sources of infection, including potential zoonotic transmission for HEV-3b (Pallerla et al., 2020).

CONCLUSION

The current study highlights the co-circulation of two distinct HEV genotypes (HEV-1a and HEV-3b) in Saudi suggesting multiple introduction pathways, including travel-related infections and potential zoonotic spillover. The detection of HEV-1a (99.7% identity with Burmese strains) indicates likely importation from South/ Southeast Asia, while HEV-3b (linked to Japanese/ Nepalese strains) raises concerns about non-endemic zoonotic or travel-associated transmission. These results underscore the necessity of targeted public health interventions, including improved diagnostic screening, zoonotic surveillance and vaccination strategies for atrisk populations. Future research should focus on expanding genomic surveillance to better understand the HEV transmission dynamics in Saudi Arabia.

Author Contributions

Conceptualization, methodology, data analysis, writing original draft and review are contributed by all authors.

Conflict of Interest

The authors declare that there is no conflict of interest.

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