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Review

The Biochemistry, Toxicology, and Uses of the Pharmacologically Active Phytochemicals: Alkaloids, Terpenes, Polyphenols, and Glycosides

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Abstract

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The biochemistry, applications, toxic potentials, and uses of alkaloids, terpenes, polyphenols, and glycosides. Traditional medicine is still recognized and accepted as the desired primary health care system in several communities, with over 80% in developing countries and about 60% of the world's population directly depending on the medicinal plants for their medical treatment and health purposes. Many parts of plants are being used to return abnormalities to normal, prevent illnesses, and/or allay symptoms. These plant parts contain bioactive compounds; terpenes, glycosides, polyphenols, and alkaloids. Alkaloids have pharmacological activities such as antiasthma, anticancer, andantimalarial. Other alkaloids possess stimulant and psychotropic activities, and are widely used as recreational drugs or in entheogenic rituals. Alkaloids can be toxic too. Although alkaloids act on diverse metabolic systems in humans and animals, they almost uniformly induce a bitter taste. The characteristics and number of the structures of phenol in polyphenols underlie the unique chemical, physical, and biological (metabolic, therapeutic, toxic, etc.) properties of specific members of the class; e.g. ellagitannin and tannic acid. Plants often contain complex mixtures of polyphenols. Several polyphenolic extracts, such as from grape seeds, olive pulp, maritime pine bark, or grape skin, are sold as ingredients for functional foods, cosmetics, and dietary supplements. Terpenes have gained public awareness by the growth and education of recreational and medical cannabis. Terpenoids and terpenes are the primary components of the essential oils of most flowery plants. Essential oils are used widely as scents and fragrances in perfumery and in traditional medicine, such as aromatherapy. Similarly, many plant glycosides are used as medicines. Amygdalin and laetrile (a synthetic derivative) were investigated as potential cancer treatment drugs; nevertheless, they are ineffective and dangerous. Pharmacologically active phytochemicals have useful applications and safety concerns.

Keywords: Medicinal Plants, Medicinal plant phytochemicals, Alkaloids, Terpenes, Polyphenols, Glycosides

INTRODUCTION

The use of plants to cure many human diseases has a long history. Several parts of plants such as the leaf, stem, bark, root, among others, are being used to prevent, allay symptoms or to revert abnormalities back to normal. As the practices of herbal remedies do not strictly adhere to facts accrued with scientific approaches, orthodox medicine perceives herbal medicines as

alternative medicines. However, most pharmaceutical products presently dispensed by physicians and pharmacists have a history of usage as herbal remedies, including digitalis, quinine, opium, and aspirin. Modern medicine today uses active compounds isolated from the higher plants, and about 80 percent of these active ingredients show positive correlation between their

traditional uses and the modern therapeutic uses (Sarkar et al., 2015).

The use of, and search for dietary supplements and drugs obtained from plants have increased recently. and researchers such as phytochemists, pharmacologists, microbiologists, and food chemists are exploring the Earth for phytochemicals and the clues that can be developed into medicines for the treatment of various diseases. Traditional medicine is still recognized and accepted as the desired primary health care system in several communities, with over 80% in developing countries and about 60% of the world's population directly depending on the medicinal plants for their medical treatment and health purposes (Shrestha and Dhillion, 2003). Most natural occurring sugar alcohols can be used for medicinal purposes, especially as laxatives (Awuchi and Echeta, 2019; Awuchi, 2017).

Currently, millions of people all-over the world consume plant based medications as part of the traditional medicine for a wide range of medical disorders. The utilization of traditional medicine in developing and underdeveloped countries directly contributes to the socio-economic status, wellbeing, and needs of the rural communities (Tabuti et al., 2003; Chiranjibi et al., 2006). People in rural areas generate income from medicinal plants, especially herbalists and traditional healers. The use of medicinal plants particularly in primary health care system has become prominent, and has led to the growing search for plants with medicinal application and use. The search and quest for plants with medicinal uses have led to many ethnobotanical studies that documented species of traditional medicinal plant, the preparation mode, and uses by local communities in many parts of the country (Tabuti et al., 2010; Lamorde et al., 2010). The identifications of these medicinal plants and in later stage the investigations of the quality and toxicity are of the utmost importance. Moreover, a lot of invaluable indigenous information about the uses of medicinal plants are being lost from one generation to the other and with the growing rate of destruction of habitats, plant resources including the medicinal plants are threatened or are getting depleted.

Phytochemicals

Alkaloids

Alkaloids are class of organic compounds that occur naturally and mostly contain basic atoms of nitrogen. Also, this group includes various related compounds with weak acidic and neutral properties. Some synthetic compounds of similar structure can also be termed alkaloids (Awuchi, 2019; Robert, 1998). Along

with carbon, hydrogen and nitrogen, alkaloids can also contain sulfur, oxygen, and, often rarely, other elements such as phosphorus, chlorine, and bromine. Morphine, the first individual alkaloid, was isolated from the opium poppy (*Papaver somniferum*) in 1804 (Andreas, 2009).

Alkaloids are produced by a wide range of organisms including fungi, bacteria, plants, and animals (Roberts, 1998). They are usually purified from the crude extracts of these organisms through acid-base extractions, or extraction followed solvent by silica-gel column chromatography (Gonçalves et al., 2013). Alkaloids have various pharmacological activities such as antimalarial (e.g. quinine), anticancer (e.g. homoharringtonine), antiasthma (e.g.ephedrine) (Kittakoop 2014), cholinomimetic (e.g. galantamine) (Russo et al., 2013), vasodilatory (e.g. vincamine), analgesic (e.g. morphine), antiarrhythmic (e.g.quinidine), antihyperglycemic activities (e.g.piperine), and antibacterial (e.g. chelerythrine). Many have found use and applications in modern or traditionalmedicine, or as the starting points for drug discovery. Many other alkaloids possess stimulant (e.g.cocaine, caffeine, theobromine, nicotine) and psychotropic activities (e.g.psilocin), and have been used as recreational drugs entheogenic rituals. Alkaloids can be toxic too (e.g.tub ocurarine, atropine) (Robbers et al., 1996). Although alkaloids act on a variety of metabolic systems in humans and in other animals, they nearly uniformly evoke a bitter taste (Awuchi, 2019).

Boundary between alkaloids and the other nitrogencontaining natural compounds is ambiguous. Compounds like proteins, amino acid peptides, nucleotides, nucleic acid, antibiotics, andamines, are usually not referred to as alkaloids. The natural compounds with nitrogen in the exocyclic position (dopamine, mescaline, serotonin, etc.) are often classified as amines than as alkaloids (Leland, 2006). Some authors and researchers, however, consider alkaloids special case of amines (Raj, 2004).

Classifications of alkaloids

There is no uniform classification. In the beginning, when there was lacking of knowledge of chemical structures, botanical classifications of the source plants was greatly relied on. Currently, this classification is considered obsolete. Compared with many other classes of natural compounds, the alkaloids are characterized by great structural diversity. More recent classification is based on the similarity of the carbon skeleton (e.g., isoquinoline-, pvridine-. indole-like) or biochemical precursor (lysine, tyrosine, tryptophan, ornithine, etc.). Nevertheless, they require compromises in the borderline cases; e.g., nicotine contains a pyrrolidine part from ornithine and a pyridine fragment from nicotinamide and as a

result can be assigned to both classes.

The alkaloids are usually divided into the following main grouping:

- a) True alkaloids; which contain nitrogen (N) in the heterocycle and have their source from amino acids. Characteristic examples arenicotine, morphine, and atropine. Also, this group includes few alkaloids that besides the nitrogen heterocycle have peptide (e.g. ergotamine) or terpene fragments (e.g., evonine). The piperidine alkaloids coniceine and coniine may be considered as true alkaloids (instead of pseudoalkaloids, as can be seen below) though they do not originate from the amino acids.
- b) Polyamine alkaloids derivatives of spermidine, spermine, and putrescine.
- c) Protoalkaloids, which contain nitrogen (not the nitrogen heterocycle) and originate from amino acids. Examples are mescaline, ephedrine, and adrenaline.
- d) Pseudoalkaloids alkaloid-like compounds which do not originate from the amino acids. This group includes steroid-like and terpene-like alkaloids, and also purine-like alkaloids such as theacrine, theophylline, caffeine, and theobromine. Some authors and researchers classify as pseudoalkaloids such compounds such as cathinone and ephedrine. Those originate from amino acid phenylalanine, but acquire their nitrogen atom through transamination, not from the amino acid.
- e) Peptide and cyclopeptide alkaloids.

Some alkaloids do not have carbon skeleton characteristic of the group they belong. Homoaporphines and galanthamine do not contain isoquinoline fragment, but they are generally attributed to the isoquinoline alkaloids.

Properties of alkaloids

Most alkaloids have oxygen in their molecular structure; the compounds are often colorless crystals at ambient conditions. The oxygen-free alkaloids, such as coniine or nicotine, are typically colorless, oily, volatile liquids. Several alkaloids are colored, like sanguinarine (orange) and berberine (yellow). Bufotenin contains an indole core and can be produced from the amino acid tryptophan in living organisms. The nicotine molecule contains both pyrrilidine and pyridine rings.

Most alkaloids are weak base, but some, such as theophylline and theobromine, are amphoteric (Spiller, 1997). Many alkaloids poorly dissolve in water but dissolve readily in organic solvents. such as chloroform, 1,2-dichloroethane, or diethyl ether. Caffeine, nicotine, cocaine, and codeine are slightly soluble in water (with solubility of ≥1g per liter), while others, including yohimbine and morphine, are very slightly soluble in water (0.1–1 g/L). Acids and alkaloids

form salts of various strengths. The salts are often freely soluble in ethanol and water and poorly soluble in many organic solvents. A few exceptions include the water-soluble quinine sulfate, and thescopol-amine hydrobromide, that is soluble in organic solvents.

Many alkaloids have a bitter taste and poisonous when consumed. Alkaloid production in plants seemed to have evolved in natural response to feeding by herbivorous animals; although, some animals have acquired the ability to detoxify alkaloids. Many alkaloids may produce developmental defects in the offsprings of animals that ingest but cannot detoxify alkaloids. A typical example is the alkaloid cyclopamine, which is produced in the leaves of the corn lily. In 1950s, about 25% of lambs born by the sheep that grazed on corn lily had severe facial deformations. These deformations ranged from deformed jaws to cyclopia. After decades of research, in 1980s, the compound 11-deoxyjervine (an alkaloid), later renamed to cyclopamine, was identified as been responsible for these deformities (Thomas et al., 2004).

Distribution of alkaloids in nature

Generally, alkaloids are generated by several living organisms, especially by the higher plants – about 10 – 25% of those contain alkaloids. Thus, earlier the term "alkaloid" was linked with plants. The alkaloids in plants is often within a few percent and is also inhomogeneous over the tissues of plants. Depending on type of plants, maximum concentration is often observed in the fruits or seeds (Strychnine tree), leaves (black henbane), bark (cinchona), or root (*Rauwolfia serpentina*). Furthermore, different tissues of same plants can contain different alkaloids.

Outside plants, alkaloids are also found in animals, such as bufotenin in the skin of many toads, in many insects, markedly ants, and in some types of fungi, such as psilocybin in fungus of the genus *Psilocybe* (Touchard *et al.*, 2016). Also, many marine organisms contain alkaloids. Several amines, such as serotonin and adrenaline, which play a significant role in the higher animals, are similar to the alkaloids in their biosynthesis and structure and are sometimes referred to as alkaloids.

Alkaloids Extraction

Due to the alkaloids structural diversity, there is no single method extracting from natural raw materials. Majority of the methods exploit the properties of most alkaloids, especially solubility in organic solvents but not in water, as well as the opposite tendency of their salts.

Many plants contain numerous alkaloids. Their mixture is first extracted and then the individual alkaloids are

separated. The plants are ground thoroughly before the extraction. Most alkaloids exist in the raw plants in form of salts of the organic acids. The extracted alkaloids can remain salts or may change into bases. The base extraction is accomplished by processing the raw materials with alkaline solutions and extracting the alkaloids bases with organic solvents, such as chloroform, diethyl ether, benzene, or 1,2-dichloroethane. The impurities are then dissolved by weak acids; which converts the alkaloid bases to salts that are washed with water. If required, an aqueous solution of the alkaloid salts is once again made alkaline and can be treated with organic solvent. The process is done repeatedly until the desirable purity is achieved.

In acidic extraction, the raw plant materials are processed by weak acidic solution (for example, acetic acid in water, methanol, or ethanol). A base is then added to transform the alkaloids to basic forms which are extracted with organic solvents (if the extraction was done with alcohol, first it is removed, and the remainder dissolved in water). In addition, the solution is purified as explained above. Alkaloids are usually separated from their mixtures using their different solubility in some solvents and different reactivity with some reagents or by distillation process.

A number of alkaloids have been identified from insects, among which the alkaloids, known as solenopsins, from fire ant venom have received greater attention from researchers (Fox, 2016). These insect alkaloids can be extracted efficiently by centrifugation of live ants or by solvent immersion of the live fire ants (Fox et al., 2018) followed by purification using silica-gel chromatography. Dosing and tracking the extracted solenopsin has been described as likely based on their absorbance peak near 232 nm (Fox et al., 2018). Crystals of piperine are extracted from the black pepper.

Biosynthesis of alkaloids

Biological precursors of many alkaloids are amino acids, such as lysine, phenylalanine, aspartic acid, anthranilic acid,tyrosine, tryptophan, ornithine, and histidine. Nicotinic acid can be synthesized from aspartic acid or tryptophan. The ways of alkaloid biosynthesis are too many and cannot be classified easily. However, there are a few distinctive reactions involved in the biosynthesis of many classes of alkaloids, including Mannich reaction and synthesis of Schiff bases.

Mannich reaction

An integral constituent of the Mannich reaction, along with a carbonyl compound and an amine, is a carbanion

that plays the role of nucleophile in the nucleophilic addition to ion formed by the reaction of the carbonyl and the amine.

The Mannich reaction can proceed both intramolecularly and intermolecularly:

$$\bigcirc \longrightarrow \bigcirc$$

Synthesis of Schiff bases

The Schiff bases can be synthesized by reacting amines with aldehydes or ketones. These reactions are common methods of producing C=N bonds.

In biosynthesis of alkaloids, such reactions can take place in a molecule, for example, in the synthesis of piperidine:

Dimer alkaloids

In addition to the above described monomeric alkaloids, there are also dimeric, trimeric and tetrameric alkaloids formed by the condensation of two, three, or four monomeric alkaloids. The dimeric alkaloids are often formed from the monomers of the same type by the following mechanisms:

- Mannich reaction, resulting in, e.g., voacamine
- Condensation of the aldehydes with amines (toxiferine)
- Michael reaction (villalstonine)
- Lactonization (carpaine).
- Oxidative addition of phenols (tubocurarine, dauricine)

There are also dimeric alkaloids produced from two distinctive monomers, such as the *vinca* alkaloids vincristine and vinblastine, which are made from the coupling of vindoline and catharanthine (Hirata *et al.*, 1994; Gansäuer *et al.*, 2007). Vinorelbine, a known novel semi-synthetic chemotherapeutic agent, is used in treatment of the non-small-cell lung cancer (Faller and Pandi, 2011). It is another derivative dimer of catharanthine and vindoline and is synthesized from anhydrovinblastine (Ngo *et al.*, 2009), starting either from leurosine (Morcillo *et al.*, 2014) or the monomers themselves.

The biological roles of alkaloids

The role of alkaloids for the living organisms that produce them is not clear. Initially it was presumed that the alkaloids are the end products of the metabolism of nitrogen in plants, as urea in mammals. Later it was shown that the concentration of alkaloids varies with time, and this hypothesis was contradicted. Also, many ants are suggested to produce alkaloids as components of their venom, however the exact pathways for the biosynthesis have not been empirically established (Pianaro et al., 2012).

Most of the functions of alkaloids already identified are related to protection. For example, liriodenine, aporphine alkaloid, produced by tulip tree protects the tree from parasitic mushrooms. Additionally, the presence of alkaloids in plant prevents chordate animals and insects from eating it. Plants use alkaloids as part of their defense mechanisms against insects, chordate animals, etc. However, some animals are adaptive to alkaloids and can use them in their own metabolism. Some alkaloid-related substances such as serotonin, histamine, and dopamine are important neurotransmitters in animals. Many alkaloids are also reported to regulate plant growth. One typical example of an organism that make use of alkaloids for protection is the Utetheisa ornatrix, commonly known as ornate moth. The pyrrolizidine alkaloids render the larvae and adult moths unpalatable to lots of their natural enemies such as insectivorous hemiptera, insectivorous bats, coccinelid beetles, and green lacewings (Conner, 2009). Another example of alkaloids in use occurs in Agonopterix alstroemeriana (the poison hemlock moth). The moth feeds on its alkaloid-rich and highly toxic host plant poison hemlock (Conium maculatum) in its larval stage. Agonopterix asltroemeriana may benefit twofold from toxicity of naturally-occurring alkaloids, both through the ability of Agonopterix alstroemeriana to recognize the Conium maculatum as the correct location for the oviposition, and through the non-palatability of the species to predators (Castells and Berenbaum, 2006). Solenopsin, a fire ant venom alkaloid, has been

shown to protect queens of the invasive fire ants during foundation of new nests, therefore playing a significant role in spread of this species of pest ant around the world (Fox *et al.*, 2019).

Applications and uses of alkaloids

Use as psychoactive drug:preparations of plants containing alkaloids and their extracts, and later the pure alkaloids. have long been applied and used as psychoactive substances. Caffeine, cathinone, and cocaine are the CNS (central nervous system) stimulants. Mescaline and various indole alkaloids (such as dimethyltryptamine, ibogaine, and psilocybin) have hallucinogenic effect. Codeine and morphine are strong narcotic pain killers.

Also, there are alkaloids which do not have strong psychoactive effects themselves, but are the precursors for semi-synthetic psychoactive drugs. e.g., pseudoephedrine and ephedrine used are to produce methamphetamine and methcathinone. Thebaine is used in synthesis of several painkillers such as oxycodone.

Use in medicine: the medical use of plants containing alkaloid has a long history, and, as a result, when the first set of alkaloids were isolated in 19th century, they instantly found application and use in clinical practice. Several alkaloids are still used in medicine, usually in the form of salts, including the following (Table 1)

Table 1. Alkaloids and their use in medicine

Alkaloid	Action
Ajmaline	antiarrhythmic
Atropine, hyoscyamine,	anticholinergic
scopolamine	
Caffeine	adenosine receptor antagonist,
	stimulant
Codeine	analgesic, antitussive
Colchicine	remedy for gout
Emetine	Emesis, antiprotozoal agent
Ergot alkaloids	Vasoconstriction, Uterotonic,
	hallucinogenic
Morphine	analgesic
Nicotine	nicotinic acetylcholine receptor
	agonist, stimulant
Physostigmine	acetylcholinesteraseinhibitor
Quinidine	Antiarrhythmic
Quinine	antimalarial, antipyretic
Reserpine	antihypertensive
Tubocurarine	Relax muscle
Vinblastine, vincristine	antitumor
Vincamine	antihypertensive, vasodilating
Yohimbine	aphrodisiac, stimulant

Many semisynthetic and synthetic drugs (medicines) are structural modifications of alkaloids, which were desi-

gned to change or enhance the primary effect of the drug and reduce undesirable side-effects. For example, an opioid receptor antagonist called naloxone is a derivative of thebaine which is present in the opium.

Use in agriculture: Prior to development of various relatively low-toxic synthetic pesticides, many alkaloids, such as salts of anabasine and nicotine, were used as insecticides. Their high toxicity to humans limited their use (György et al., 2002).

Polyphenols

Polyphenols also called polyhydroxyphenols structural class of mostly natural, but also semisynthetic synthetic, organic chemicals characterized presence of huge multiples of phenol structural units 2011; (Quideau al., Quideau, 2011). The et characteristics and number of the structures of these phenol underlie the unique chemical, physical, and biological (therapeutic, metabolic, toxic, etc.) properties of specific members of the class. Some examples are ellagitannin and tannic acid (tannin). The historically significant chemical class of the tannins is subset of the polyphenols (Quideau et al., 2011). Plant-derived polyphenol, tannic acid (tannin), is formed by the esterification of ten equivalents of phenylpropanoidderived gallic acid to glucose (a monosaccharide) core from primary metabolism. Phenol-phenolate resonance structures and equilibrium giving rise to aromatic reactivity.

Individual polyphenols engage in the reactions associated to both their core phenol structures, their linkages, and the types of glycosides they form. The standard phenolic reactions are oxidations to para- and ortho-auinones (which contributes to antioxidant characteristics), underlying aromatic transformations related to presence of the phenolic hydroxyl, and ionization (which contributes to complexation and solubility); reactions related to their linkages include oxidative and hydrolytic bond cleavages, and nucleophilic additions. Additionally, a traditional feature of the polyphenols was their ability to form specific, characteristic metal complexes (Haslam and Cai, 1994).

Chemical structure and synthesis of polyphenols

Structural features of polyphenols

Opposite to the smaller phenols, the polyphenols are often macromolecules (larger molecules) deposited in the cell vacuoles. The upper limit of molecular weight for small molecules is 800 Daltons, which makes allowance for the possibility of rapid diffusion across cell membranes so they can reach the intracellular sites of

action or can remain as pigments once senesces. Hence, numerous larger polyphenols are biosynthesized in-situ from the smaller polyphenols to non-hydrolysable tannins and remain undiscovered and unidentified in the plant matrix. Most polyphenols have repeating phenolic moieties of resorcinol, pyrogallol, phloroglucinol, and pyrocatechol linked by more stable C-C bonds (non-hydrolysable condensed tannins) or esters (hydrolysable tannins). Proanthocyanidins are polymeric units of epicatechin and catechin. Resorcinol (1,3-benzenediol) and catechol types of polyphenols have two, and phloroglucinol (benzenetriol) pyrogallol types have three phenol hydroxyl groups, respectively, although mixing of these types in polyphenols is also likely. The phenolic substructures arise from many biosynthetic pathways (WBSSH definition), especially polyketide and phenylpropanoid branches aimed at plant and the related secondary metabolites. The C-glucoside substructure of polyphenols is typified by puerarin (the phenol-saccharide conjugate). a mid-molecular-weight natural plant product. The phenol attachment to the saccharide is through a carbon-carbon bond (C-C). The isoflavone and benzopyran (10-atom "fused ring" system of isoflavone), also a structural feature here, is commonly found in polyphenols.

Other than hydroxyl groups, polyphenols have heteroatom substituents; ester and ether linkages are common, as are many carboxylic acid derivatives; ester linkages are common in the hydrolysable tannins. In addition to the simple heteroatom links, the carbon frameworks may become complex, e.g., several C-C linkages join hydrolytically labile ethers and esters as common in non-hydrolysable tannins. The biaryl/biphenyl substructure of polyphenols can be prepared by synthesis using the copper-mediated Ullmann reaction. Also, the C-C bond in biaryls in nature is synthesized through a metal-mediated coupling reaction, always involving iron. The biaryl substructure is found in ellagic acid. Because of the preponderance of the saccharidederived core structures, as well as spiro- and other structure types, natural chiral (stereo) centers abound.

Chemical synthesis of polyphenols

True polyphenols from tannin and other WBSSH types are consistently biosynthesized in the natural sources where they derive; their chemical syntheses (with standard bench organic chemical methods) were to some extent limited until first decade of the new millennium due to these syntheses involve challenging stereoselectivity and regioselectivity issues (Krohn *et al.*, 2010). Early work focused on achiral synthesis of phenolic-related constituents of polyphenols in the late 1970's, and the Nelson and Meyers synthesis of permethyled derivative of the ubiquitous and abundant

diphenic acid core of the ellagitannins in 1994 (Nelson and Meyers, 1994) followed by the stereoselective synthesis of the more complex permethylated structures such as Itoh and coworker's synthesis of permethylated pedunculagin with specific attention to the axial symmetry issues in 1996, and Lipshutz and coworkers' (+)tellimagrandin II derivative in 1994 (Itoh et al., 1996). The total synthesis of a completely unmasked polyphenol that of ellagitannin tellimagrandin I, was diastereoselective sequence reported by Feldman, Ensel and Minard 1994. Further total syntheses of the in deprotected polyphenols that followed were headed by the Feldman group, for example in Feldman and Lawlor's synthesis of ellagitannin, coriariin A and other tannin relations (Feldman et al., 2000). Khanbabaee and Grosser achieved a relatively efficient total synthesis of the pedunculagin in 2003 (Khanbabaee and Grosser, 2003; Feldman, 2004).

Work proceeded with a focus on enantioselective total syntheses, for example, on atroposelective syntheses of the axially chiral biaryl polyphenols (Bringmann et al., 2011; Pouysegu et al., 2011), with recent further significant work including the controlled assembly of various polyphenols according to integrated strategies, in the syntheses of extended oligomeric catechins) of procyanidins (an many groups and of resveratrol polyphenols by Snyder group at Columbia which included the diverse ampelopsins G and H, carasiphenols B and C, and nepalensinol B (Snyder et al., 2011). A biomimetic synthesis, as well as the first formal total synthesis of the 5-O-Desgallovl-epipunicacortein A, an additional ellagitannin in its Cglucosyl (the C-glucoside subclass), has recently been accomplished also. The novel strategies and methods mentioned in these recent examples helped in opening the field of the polyphenol chemical synthesis to such an unprecedented degree (Quideau, 2011).

Chemical properties and uses of polyphenols

Chemical properties of polyphenols

Polyphenols are molecules owing their Ultraviolet/ Visible absorptivity to aromatic structures with great conjugated systems of pi electron configurations; also, they have autofluorescence properties, particularly lignin and the phenolic part of the suberin. They are reactive species to oxidation. ABTS may be used to characterize polyphenol oxidation products (Osman *et al.*, 2006). Polyphenols also characteristically possess an important binding affinity for proteins, which may lead to the formation of the soluble and the insoluble protein-polyphenol complexes (Papadopoulou and Frazier, 2004).

Chemical uses of polyphenols

Traditionally, some polyphenols are used as dyes. For example, in the Indian subcontinent, the peel of pomegranate, high in tannins and other polyphenols, or the pomegranate juice, is employed in dyeing of nonsynthetic fabrics (Jindal and Sharma, 2004). Polyphenols, especially tannins, were traditionally used for tanning leather and are currently used as precursors in green chemistry especially to produce plastics or resins by polymerization with or without use of formaldehyde or the adhesives for particleboards. Generally, the objectives are to make use of pecan shells left after processing or the plant residues from grape, olive (known as pomaces) (Aizpurua-Olaizola *et al.*, 2015). Pyrocatechin and pyrogallol are among oldest photographic developers.

Polyphenols Biology

The biological role in plants

Both the larger polyphenols play important roles in the ecology of many plants (Awuchi, 2019). Their known effects in the tissues of plants can be divided into following categories (Lattanzio *et al.*, 2006):

- UV screens to provide coloration (plant pigments) and to protect against ionizing radiation.
- Signaling molecules in growth processes, including ripening.
- Release and suppression of the growth hormones such as auxin.
- Prevention of microbial infections (phytoalexins).
- Deterrence of herbivores (sensory properties).

Polyphenols occurrence in nature

The most abundant polyphenols in nature are the condensed tannins, which are found in virtually all plants families. Larger polyphenols are always concentrated in bark layers, flowers, fruits, the leaf tissue, and the epidermis but also play significant roles in nutrient cycles in forest ecology, and the decomposition of forest litter. Absolute concentrations of the total phenols in plant tissues widely differ depending on the type of polyphenols, assay, and literature source; they range from 1 to 25% total polyphenols and natural phenols, calculated with reference to dry green leaf mass (Hättenschwiler and Vitousek, 2000).

High levels of polyphenols in various woods can explain their natural preservations against rot. Flax and a submerged aquatic plant called *Myriophyllum spicatum* secrete polyphenols which are involved in allelopathic interactions (Popa *et al.*, 2008). Also, poly-

phenols are found in animals. In some arthropods such as crustaceans and insects polyphenols play a role in sclerotization (epicuticle hardening). The of hardening is due the presence the to enzyme polyphenol oxidase. Apparently, there is no polyphenol tanning occurring in the arachnids' cuticle. In crustaceans, there is second oxidase activity resulting in cuticle pigmentation.

Biosynthesis and metabolism of polyphenols

Polyphenols incorporate smaller parts (components) and building blocks from the simpler natural phenols that originate from the phenyl propanoid pathway for phenolic acids or shikimic acid pathway for the gallotannins and analogs. Caffeic acid derivatives and flavonoids are biosynthesized from malonyl-CoA and phenyl alanine. Complex gallotannins develop through dimerization processes resulting in hydrolyzable tannins or through invitro oxidation of 1,2,3,4,6-pentagalloyl-glucose. For anthocyanidins, the precursors of condensed tannin biosynthesis, leucoanthocyanidin reductase (LAR) and dihydroflavonol reductase are crucial enzymes with the subsequent addition of epicatechin and catechin moieties for larger, non-hydrolysable tannins (Tanner et al., 2003). The glycosylated form develops from the glucosyltransferase activity and increases the polyphenols solubility (Krasnow and Murphy, 2004).

Polyphenol oxidase (PPO) is an enzyme which catalyzes the oxidation of o-diphenols to give o-quinones. It is the rapid polymerization of o-quinones to produce red, black, or brown polyphenolic pigments that is responsible for browning in fruits, such as browning in apple when cut or bruised. In insects, polyphenol oxidase serves for the hardening of the cuticle. Laccase is a major enzyme which initiates the cleavage of hydrocarbon rings that catalyze the addition of a hydroxyl group (-OH) to phenolic compounds. The enzyme can be found in fungi such as Panellus stipticus, organisms that break down the lignin, a complex polymer (aromatic) in the wood that is highly resistant to the degradation by conventional and normal enzyme systems. Hypericin, phenolic lipids, and anthracyclines are derived from polyketides cyclisation (Hertweck, 2009).

Polyphenols content in food

Generally foods contain complex polyphenols mixtures. The most important food sources are the commodities commonly consumed in large quantities such as green tea, black tea, fruit and vegetables, red wine, coffee, extra virgin olive oil, chocolate, and olives. Herbs and spices, algae and nuts are also potentially

significant for supplying some polyphenols. Many polyphenols are specific to particular foods (phloridzin in apples, isoflavones in soya, flavanones in citrus fruit); while others, such as quercetin, are detected in all the plant products such as vegetables, cereals, leguminous plants, fruits, tea, and wine (d'Archivio *et al.*, 2010).

Some polyphenols are considered anti-nutrients, substances that interfere with absorption of essential nutrients, particularly iron and other metal ions, and also by binding to the digestive enzymes and other proteins, especially in ruminants (Mennen *et al.*, 2005). However, a few polyphenols have antioxidant properties. The phenolic and the carotenoid compounds with antioxidant properties in the vegetables have been found to be significantly retained better by steaming than by frying. Polyphenols in beer, wine, and numerous nonalcoholic juice beverages could be removed by using finings, substances that are often added near or at the completion of the brewing processing.

Potential health effects of polyphenols

Many polyphenolic extracts from sources, such as grape seeds, grape skin, maritime pine bark, and olive pulp, are sold as ingredients in cosmetics, functional foods, and dietary supplements with little or no proof of effect or legal health claims (Gross, 2009). Some polyphenol ingredients have self-aclaimed GRAS status in the US.

The diverse structures and indeterminate metabolic fate of the phenolic compounds following digestion prevent the understanding about their potential health effects. Especially, due to the presumed antioxidant role of the polyphenols in vivo cannot be well established, the US FDA issued labeling direction to manufacturers that the polyphenols cannot be declared as antioxidant nutrients except physiological evidence exists to confirm such a qualification and also a DRI value has been established (Gross, 2009; US Food and Administration, 2017). Furthermore, since the purported health claims for some specific polyphenol-enriched foods remain unproven (Halliwell, 2007), health statements regarding the polyphenols on product labels are banned by the EFSA and the FDA(EFSA, 2011). However, the EFSA recently recognized health claims of some specific products, such as olive oil and cocoa (EFSA, 2014).

Compared with the *in vitro* effects of polyphenols, the possible *in vivo* functions remain unknown due to (a) long-term studies failing to show effects with an action mechanism, sensitivity and efficacy or specificity (Gross, 2009); (b) the absence of any validated *in vivo* biomarkers; and (c) invalid applications of high, non-physiological test concentrations in *in vitro* studies, which are subsequently not relevant for design of *in vivo* experiments.

Use of polyphenols in traditional medicine

Herbal teas contain soluble polyphenols that impart astringent effects believed to have medicinal properties (Haslam *et al.*, 1989). In Ayurveda system of folk medicine, pomegranate and its polyphenolrich peel are thought to be useful for therapeutic purposes.

Sensory properties of polyphenols

With respect to food and beverages, the main cause of astringency is not entirely understood, but it is biochemically measured as the ability of a substance or compound to precipitate proteins. A review article published in 2005 found that bitterness decreases and astringency increases with mean degree of polymerization. For the water soluble polyphenols, the molecular weights between 500 - 3000 were reported to be necessary for protein precipitation. However, the smaller molecules might still have some astringent qualities possibly due to formation of non-precipitated complexes with proteins or the cross-linking of proteins with the simple phenols that have 1,2,3-trihydroxy or 1,2dihydroxy groups (Lesschaeve and Noble, 2005). Also, flavonoid configurations can cause significant differences in sensory properties, for example, epicatechin is more astringent and bitter than its chiral isomer catechin. By contrast, hydroxycinnamic acids do not have any astringent qualities, but are bitter (Hufnagel and Hofmann, 2008).

Extraction of polyphenols

Extraction of polyphenols can be done using a solvent like methanol, water, hot water, methanol/formic acid, water/methanol/acetic or formic acid etc. The extraction by liquid—liquid or countercurrent chromatography can also be done. Solid phase extraction can also be done on C18 sorbent cartridges. Some other techniques are heat reflux extraction, ultrasonic extraction, microwave-assisted extraction (Pan, 2003), use of ethanol in immersion extractor, orcritical carbon dioxide, pressurized liquid extraction. The extraction conditions (temperature, solvent and concentrations, ratio of solvent to raw material, extraction time) have to be optimized.

Mainly found in the fruit seeds and skins, high levels of polyphenols may only reflect the EPP (measured extractable polyphenol) content of a fruit which also may contain non-extractable polyphenols. The black tea contains high quantities of polyphenol which makes up 20% of its weight (Arranz et al., 2009). Purification can be accomplished by preparative chromatography. Concentration can be done by ultrafiltration.

Analytical techniques

The analysis techniques are those of the phytochemistry: including extraction, isolation, structural elucidation, and then quantification (Owen *et al.*, 2003). Phosphomolybdic acid is used as reagent for staining phenolics in TLC (thin layer chromatography). Polyphenols may be studied by spectroscopy, particularly within the ultraviolet domain, by paper or fractionation chromatography. They can also be analyzed by chemical characterization.

Instrumental chemistry analyses include separation by HPLC (high performance liquid chromatography), and especially by RPLC (reversed-phase liquid chromatography), can be coupled to mass spectrometry (MS). Purified compounds can be identified by means of nuclear magnetic resonance (NMR). The DMACA reagent is a histological dye specific to the polyphenols used in microscopy analyses. Also, auto-fluorescence of polyphenols can be used, especially for localization of suberin and lignin.

Polyphenol content can be auantified isolation/separation by volumetric titration. Perman ganate, an oxidizing agent, is used to oxidize known concentration of a standard solution of tannin, producing a standard curve. Tannin content of the unknown (sample) is then expressed as equivalents of appropriate condensed or hydrolysable tannin. Some methods for the quantification of total content of polyphenol are based on colorimetric measurements. Various tests are relatively specific to polyphenols (e.g. the Porter's assay). The total phenols (or the antioxidant effect) can be measured with the Folin-Ciocalteu reaction. The results are typically expressed as the gallic acid equivalents. polyphenols are rarely evaluated by antibody technologies.

Some other tests measure a fraction's antioxidant capacity. Some make use of ABTS radical cation which is very reactive to most antioxidants including thiols, phenolics, and vitamin C (Walker and Everette, 2009). During this reaction, blue ABTS radical cation is transformed back to its initial colorless neutral form. The reaction can be spectrophotometrically monitored. This assay is often called the TEAC (Trolox equivalent antioxidant capacity) assay. The reactivity of the numerous tested antioxidants are always compared to that of Trolox, a vitamin E analog.

The other antioxidant capacity assays that use Trolox as standard include oxygen radical absorbance capacity (ORAC), diphenylpicrylhydrazyl (DPPH), ferric reducing ability of plasma (FRAP) (Pulido *et al.*, 2000) assays or inhibition of the copper-catalyzed *in vitro* human oxidation of low-density lipoprotein. New methods including use of biosensors can assist in monitoring the content of polyphenols in food (Mello *et al.*, 2003). Quantitative results produced by the mean

of the diode array detector-coupled high performance liquid chromatography are in general given as relative rather than the absolute values as there is lack of commercially available standards for entire polyphenol molecules.

Terpenes

Terpenes are large, diverse class of organic а compounds, made by various plants, especially conifers, and by some insects (Eberhard, 2006; Davis and Croteau, 2000). Terpenes are hydrocarbons. They often have strong odour and may protect plants that produce them by attracting predators and the parasites of herbivores, and by deterring herbivores (Martin et al., 2003; Pichersky, 2006). Although occasionally used interchangeably with the terpenes the terpenoids (or isoprenoids) are the modified terpenes as they have additional functional groups. often oxygencontaining.

Terpenes are the major constituents of turpentine and of rosin produced from resin. Terpenes are also main biosynthetic building blocks. For example, steroids are derivatives of triterpene squalene. Terpenes and terpenoids are the primary components of the essential oils of various types of flowers and plants. Essential oils are widely used as fragrances in traditional medicine and perfumery, such as aromatherapy. Vitamin A is a terpenoid. The synthetic variations and the derivatives of natural terpenes and terpenoids greatly expand the range of flavors used in food additives and aromas used in perfumery.

Structure and biosynthesis

Terpenes are biosynthetically derived from units of isopentenyl pyrophosphate. Although the structures of terpenoids are reorganized as derivatives of isoprene (i.e. 2-methyl-1,3-butadiene), the isoprene is not involved in the terpenoids biosynthesis. The C_5 rule or the biogenetic isoprene rule was described by Leopold Ružička (in 1953), who explained that terpinoids can be pictured as the result of linking units of isoprene head-to-tail to form chains that can be arranged to give rings.

There are two metabolic pathways which create terpenoids: the Mevalonic acid pathway and the 2-*C*-methyl-D-erythritol 4-phosphate/1-deoxy-D-xylulose 5-phosphate (MEP/DOXP) pathway.

Many organisms produce terpenoids through the pathway of HMG-CoA reductase, called the Mevalonate pathway, which produces cholesterol too. One of the intermediate compounds and substances in this pathway is the mevalonic acid. The range of reactions take place

in cytosol. The MEP/DOXP pathway, also called mevalonic acid-independent pathway or non-mevalonate pathway, takes place in the apicomplexan protozoa and plastids of plants, as well as in several bacteria.

Geranyl pyrophosphate is converted to geranylgeranyl pyrophosphate and farnesyl pyrophosphate, respectively C20 and C15 precursors to diterpenes and sesquiterpenes (as well as diterpenoids and sesequiterpenoids). The biosynthesis is mediated by terpene synthase (Kumari *et al.*, 2017; Pazouki and Niinemets, 2016).

Types of terpenes

Terpenes may be classified according to the number of units of isoprene in the molecule; prefix in the name specifies the number of terpene units required to assemble the molecule.

- a) Hemiterpenes are made of a single isoprene unit. Isoprene is considered the only hemiterpene; oxygencontaining derivatives such as isovaleric acid and prenol are hemiterpenoids.
- b) Monoterpenes consist of *two* units of *isoprene* and have the general molecular formula $C_{10}H_{16}$. The examples of monoterpenes and monoterpenoids are terpineol (in lilacs), geraniol, limonene (in citrus fruits), linalool (in lavender), pinene (in pine trees), and myrcene (in hops), (Breitmaier, 2006).
- c) Sesquiterpenes (The *sesqui* prefix means one and a half) consist of *three* units of *isoprene* and have the general molecular formula $C_{15}H_{24}$. The examples of sesquiterpenes and sesquiterpenoids are farnesol, humulene, farnesenes, etc.
- d) Diterpenes consist of *four* units of *isoprene* and have the general molecular formula $C_{20}H_{32}$. They are derived from geranylgeranyl pyrophosphate. Typical examples of diterpenes and diterpenoids are kahweol, cembrene, taxadiene (precursor of taxol), and cafestol. Diterpenes also form basis for important biological compounds such as phytol, retinol, and retinal.
- e) Sesterterpenes are terpenes having 25 carbons and *five* units of *isoprene* and are rare relative to the other sizes. An example of sesterterpenoid is geranylfarnesol. The *sester* prefix signifies two and a half.
- f) Triterpenes have the molecular formula $C_{30}H_{48}$ and consist of six isoprene units. The linear triterpene squalene, major shark liver oil constituent, is derived from reductive coupling of two farnesyl pyrophosphate molecules. Squalene is then biosynthetically processed to generate either cycloartenol or lanosterol, the structural precursors to all steroids.
- g) Sesquarterpenes have the molecular formula $C_{35}H_{56}$ and are composed of *seven isoprene* units. Sesquarter

penes are typically microbial in origin. Examples of sesquarterpenoids are tetraprenylcurcumene and ferrucicadiol.

- h) Tetraterpenes contain *eight* units of *isoprene* and have the general molecular formula $C_{40}H_{64}$. Important biological tetraterpenoids include the monocyclic gammacarotene, the bicyclic beta- and alpha-carotenes, and the acyclic lycopene.
- i) Polyterpenes consist of long chains of *several isoprene* units. Some plants produce a polyisoprene with the *trans* double bonds, called gutta-percha. Natural rubber consists of polyisoprene with the *cis* double bonds.
- j) Norisoprenoids, such as C_{13} -norisoprenoids 3-oxo- α -ionol found in Muscat of Alexandria leaves and the 7,8-dihydroionone derivatives, such as 3-oxo-7,8-dihydro- α -ionol and megastigmane-3,9-diol found in wine (responsible for some spice notes in Chardonnay) or the Shiraz leaves (both grapes in species *Vitis vinifera*), can be produced by fungal peroxidases (Zelena *et al.*, 2009) or glycosidases.

Properties and uses of terpenes

Terpenes have many desirable properties for use in pharmaceutical, food, cosmetics, biotechnology industries (Thimmappa et al., 2014). Terpenes are valuable active ingredients as part of the natural agricultural pesticides. They are used by the termites of subfamily Nasutitermitinae to ward off predators such as insects, specialized mechanism through use of as "fontanellar gun". Higher quantities of terpenes are released from trees in warmer weather, and can be a natural form of the cloud seeding. The cloud reflects sunlight, allowing forest temperature to regulate (Adam, 2008). The flavor and aroma of hops comes partly from sesquiterpenes (mainly β-caryophyllene and humulene), which affect beer quality (Steenackers et al., 2015). The genomes of species of 17 plants contain genes that encode the terpenoid synthase enzymes imparting the terpenes with their basic structure, and the cytochrome P450s which modify this basic structure (Boutanaev et al., 2015). Terpenes are also major components of Cannabis sativa plants, which have at least 120 identified compounds (André et al., 2016; Booth et al., 2017). Terpenes have gained public awareness by the growth and education of recreational and medical cannabis. Companies and organizations operating in the cannabis markets have pushed marketing and education of terpenes in their products as a major way to differentiate the taste and effects of cannabis. Entourage Effect, which explains the synergy of terpenes, cannabinoids, and other compounds in plants, has also helped create further awareness and raise the demand for terpenes in the products of cannabis.

Industrial syntheses

While terpenes and terpenoids widely occur, their extraction from natural sources is always challenging. Consequently. thev are often produced petrochemicals by chemical synthesis. In one route, acetylene and acetone are condensed to yield 2-Methylbut-3-yn-2-ol, which is then extended with the acetoacetic ester to yield geranyl alcohol. Many others are prepared from those terpenoids and terpenes which are readily isolated in amount, say from the paper and the tall oil industries. For instance, α-pinene, which is readily obtained from natural sources, is converted to camphor and citronellal. Citronellal is also converted to menthol and rose oxide (Eberhard, 2006).

Glycosides

A glycoside is a molecule with a sugar bound to another functional group through a glycosidic Glycosides play numerous important roles in the living organisms. Several plants store chemicals in form of glycosides, which mav be by enzyme hydrolysis (Brito-Arias, 2007), causing the sugar part to break off, resulting in the availability of chemical for use. Lots of such plant glycosides are used as medicines. Many species of Heliconius butterfly have the capability to incorporate these plant compounds as form of chemical defense against their predators. In humans and animals, poisons are often bound to molecules of sugar as part of their elimination mechanisms from the body. Amygdalin the first glycoside ever identified, by the French chemists Antoine Boutron-Charlard and Pierre Robiquet, in 1830.

Any molecule with a sugar group bonded via its anomeric carbon to another group through a glycosidic bond is a glycoside. Glycosides can be linked by an S-(a thioglycoside), C- (a C-glycoside), O- (an O-glycoside), or N- (a glycosylamine) glycosidic bond. According to IUPAC, the preferred term is the C-glycosyl compound; the name C-glycoside is a misnomer (IUPAC, 2009). The given definition is used by IUPAC, which recommends the Haworth projection to appropriately assign stereochemical configurations (Lindhorst, 2007). Many authors require in addition that sugar be bonded to non-sugar for a molecule to qualify as a glycoside, thereby excluding polysaccharides. Then the sugar group is known as the *glycone* and the non-sugar group known as the *genin* or *aglycone* part of the glycoside. The glycone may consist of monosaccharide (a single sugar group), disaccharide (two sugar groups), or oligosaccharide (several sugar groups).

Gycosides related compounds

Molecules containing N-glycosidic bond are called glycosylamines. Many biochemistry authors call these compounds *N-glycosides* and place them in the group of glycosides; which is considered a misnomer and strictly discouraged by the IUPAC. Glycosylamines and glycosides are together grouped as glycoconjugates; other glycoconjugates include peptidoglycans, glycolipids, lipopolysaccharides, glycoproteins, and glycopeptides.

Chemistry of glycosides

The glycone and aglycone parts can be separated chemically by hydrolysis in presence of acid and can also be hydrolyzed by alkali. Also, there are many enzymes that form and break glycosidic bonds. Most significant synthetic enzymes in nature are the glycosyltransferases, and the most significant cleavage enzymes are glycoside hydrolases. Glycosynthases (genetically altered enzymes termed) have been developed to form glycosidic bonds in outstanding yield.

There are many ways glycosidic bonds can be chemically synthesized. The Koenigs-Knorr reaction involves the condensation of glycosyl halides and the alcohols in presence of metal salts such as mercuric oxide or silver carbonate. Fischer glycosidation refers to synthesis of glycosides by reaction of unprotected monosaccharides with the alcohols (often as solvent) in presence of strong acid catalyst.

Classification of glycosides

Glycosides can be classified by the type of glycosidic bond, by the glycone, and by the aglycone.

By type of glycosidic bond

Dependent on whether the glycosidic bond lies above or below the plane of the cyclic molecule of sugar, glycosides are classified as β -glycosides or α -glycosides. Some enzymes such as the α -amylase can hydrolyze α -linkages alone; others, such as the emulsin, can only affect β -linkages.

The Four type of linkages present between the glycone and the aglycone are:

- N-linkage/glycosidic bond
- O-linkage/glycosidic bond
- C-linkage/glycosidic bond, nonhydrolysable by enzymes or acids
- S-linkage/glycosidic bond

By glycone (presence of sugar)

If glycone group of any glycoside is glucose, the molecule is referred to as a glucoside; if it is fructose, the molecule is referred to as a fructoside; if it is glucuronic acid, the molecule is referred to as a glucuronide; etc. In the body, toxic substances are usually bonded to glucuronic acid to increase their solubility in water; then the resulting glucuronides are excreted.

By aglycone

Glycosides can also be classified according to the aglycone chemical nature. For purposes of pharmacology, food chemistry, and biochemistry, this is the major useful classification.

Alcoholic glycosides; Example of alcoholic glycoside is salicin found in the genus *Salix*. In the body, salicin can be converted into salicylic acid, which is closely related to the aspirin and hasantipyretic, antiinflammatory, and analgesic effects.

Steviol glycosides; these glycosides have the steviol as the aglycone part. Steviol glycosides are sweet glycosides in the stevia plant Stevia rebaudiana Bertoni. They have 40 to 300 times the sweetness of sucrose. Stevioside and rebaudioside A, the two primary glycosides, are used as natural sweeteners in some countries. Glucose or combinations of rhamnose-glucose are bound to the aglycone ends to form the different compounds.

Anthraquinone glycosides; these group of glycosides contain an aglycone group which is a derivative of anthraquinone. Anthraquinone glycosides have a laxative effect, and are mainly found in the dicot plants except the Liliaceae family which are monocots. Anthranol and anthron are reduced forms of anthraquinone. Anthraquinone glycosides are present in rhubarb, senna, and Aloe species.

Cardiac glycosides or steroidal glycosides; here the aglycone part is steroidal nucleus. They are found in the plant genera *Scilla*, *Strophanthus*, and *Digitalis*. They are used in treatment of heart diseases, for example, congestive heart failure (historically as now recognized does not improve survivability; other agents are currently preferred) and arrhythmia.

Thioglycosides; these compounds contain sulfur; as the name implies (q.v. thio-). Examples are sinalbin, found in white mustard, and sinigrin, found in black mustard

Iridoid glycosides; these contain iridoid group; e.g. theviridoside, Loganin, aucubin, Geniposidic acid, and Catalpol.

Coumarin glycosides; the aglycone is either coumarin or a derivative. Example is apterin which has been shown

to dilate the coronary arteries and block calcium channels. Some coumarin glycosides are obtained from the dried leaves of the *Psoralea corylifolia*.

Chromone glycosides; the aglycone, in this case, is called benzo-gamma-pyrone.

Cyanogenic glycosides; in cyanogenic glycosides, the aglycone contains a cyanohydrin group. The plants that make the cyanogenic glycosides store them in the plant vacuole, but, if the plant is under attack, they are released and activated by enzymes in cytoplasm. These do away with the sugar part of the molecule, causing the cyanohydrin structure to collapse, resulting in the release of toxic hydrogen cyanide (HCN). Storing them in their inactive forms in the plant vacuole prevents them from damaging and harming the plant under normal conditions (Gleadow and Møller, 2014).

Along with playing an important role in deterring herbivores, in many plants they control carbon and nitrogen transport, germination, bud formation, and possibly act as antioxidants (Gleadow and Møller, 2014). The making of cyanogenic glycosides is a conserved function of evolutionary, appearing in the species as old as the ferns and as recent as the angiosperms (Gleadow and Møller, 2014). These compounds are produced by about 3,000 species; in the screens they are found in approximately 11 percent of cultivated plants but only 5 percent of plants overall—humans appear to have selected for them (Gleadow and Møller, 2014).

Examples include prunasin and amygdalin which are produced by the bitter almond tree; other plant species that produce cyanogenic glycosides are barley, flax, sorghum (in which dhurrin, first cyanogenic glycoside to officially identified, was first isolated), white clover (Gleadow and Møller, 2014), and cassava (which makes linamarin, iso-linamarin, and lotaustralin). Amygdalin and a synthetic derivative known as laetrile were examined as potential drugs for cancer treatment and were promoted heavily as alternative medicine; however, they are ineffective and dangerous (Milazzo and Horneber, 2015). Some species of butterfly, such as Parnassius smintheus and Drvas iulia. have evolved to use cyanogenic glycosides in their host plants for protection against predators via their unpalatability.

Flavonoid glycosides; the aglycone in the flavonoid glycosides is a flavonoid. The examples of this big group of glycosides include:

- Naringin (glycone: Rutinose, aglycone: Naringenin)
- Hesperidin (glycone: Rutinose, aglycone: Hesperetin)
- Quercitrin (aglycone: Quercetin, glycone: Rhamnose)
- Rutin (aglycone: Quercetin, glycone: Rutinose)

They are also known to reduce capillary fragility. Among the important flavonoids effects are their antioxidant effect.

Phenolic glycosides; in this group, the aglycone is simple phenolic structure. Example is arbutin in Common Bearberry Arctostaphylos uva-ursi, which has urinary antiseptic effects.

Saponins: saponin glycosides are found in liquorice. They also cause the hemolysis of red blood cells. When shaken with water, the compounds give a permanent froth. Their medicinal value is due to their antiinflammatory, expectorant, and corticoid effects. Steroid saponins. in the Dioscorea wild e.g., vamsapogenin diosgenin—in the form of its alycoside dioscin—is an essential starting material for the production of semisynthetic glucocorticoids and many other steroid hormones such as progesterone. Ginsenosides are the triterpene glycosides and the Ginseng saponins from Panax Ginseng A. C. Meyer, (Chinese ginseng) and the Panax quinquefolius (American Ginseng). Generally, in organic chemistry, the use of the word "saponin" is discouraged, because several plant components can produce foam. and various triterpene-glycosides are amphipolar in certain conditions, acting as a surfactant. Modern uses of saponins in biotechnological industries are as adjuvants in vaccines: the Quil A and its deriv-ative QS-21 which are isolated from the bark of the Quillaja saponaria Molina, to stimulate both Th1 immune response and production of CTLs (cytotoxic T-lymphocytes) against the exogenous antigens make them ideal for application and use in subunit vaccines and in the vaccines directed against intracellular pathogens and also for therapeutic cancer vaccines though with aforeside-effect of hemolysis mentioned (Sun et 2009). Also, saponins are natural ruminal antiprotozoal agents and are potential to improving the ruminal microbial fermentation, thus reducing methane production and ammonia concentrations in ruminant animals (Patra and Saxena, 2009).

CONCLUSION

Various parts of plants such as leaf, stem, bark, root, etc. are being used to prevent, allay symptoms or revert abnormalities back to normal. These plants contains bioactive compounds such as terpenes, glycosides, polyphenols, and alkaloids. Alkaloids have several pharmacological activities including antimalarial ephedrine), quinine), antiasthma (e.g. anticancer (e.g. homoharringtonine). Other possess alkaloids psychotropic (for example psilocin) and stimulant activities (e.g. caffeine, nicotine, cocaine, theobro-mine), and are used in entheogenic rituals and as recreational drugs. Many alkaloids are toxic too (e.g. tubocurarine, atropine). Although alkaloids act on many metabolic systems in humans and animals, they almost uniformly evoke a bitter taste. The number and characteristics of the phenol

structures in polyphenols underlie the unique chemical, physical, and biological (metabolic, therapeutic, toxic, etc.) properties of some members of the class; examples include tannic acid and ellagitannin. In general, foods contain complex mixtures of polyphenols. Several polyphenolic extracts, including from grape seeds, grape skin, maritime pine bark or olive pulp, are sold as ingredients in dietary supplements, functional foods, and cosmetics. Terpenes and terpenoids are primary constituents of most essential oils of many plants. Essential oils are widely used as fragrances in traditional medicine and perfumery, such as aromatherapy. Many plant glycosides are regularly used as medications. Amygdalin and a synthetic derivative, known as laetrile, were investigated as potential drugs to treat cancer and were heavily promoted as alternative medicine; however, they are ineffective and dangerous.

REFERENCES

- Adam D (2008). "Scientists discover the cloud-thickening chemicals in trees that could offer new weapon in fight against global warming". The Guardian.
- Aizpurua-Olaizola, O; Ormazabal, Markel; Vallejo, A; Olivares, Maitane; Navarro, Patricia; Etxebarria, N; and Usobiaga, A (2015).Optimization of Supercritical the Fluid Consecutive Extractions of Fatty Acids & Polyphenols from Vitis Vinifera Grape Wastes. *J Food Sci.* 80 (1): E101–7. doi:10.1111/1750-3841.12715.
- André MC, Hausman FJ, Guerriero G (2016). "Cannabis sativa: The Plant of Thousand and One Molecules". *Frontiers in Plant Science*. 7: 19. doi:10.3389/fpls.2016.00019.
- Arranz, Sara; Saura-Calixto, Fulgencio; Shaha, Shika; and Kroon, P. A. (2009). "High Contents of the Nonextractable Polyphenols in Fruits Suggest Polyphenol Contents of Plant Food Have Been Underestimated". *J Agric and Food Chem.* 57 (16): 7298–7303. doi:10.1021/jf9016652.
- Awuchi CG (2019). Medicinal Plants: the Medical, Food, & Nutritional Biochemistry & Uses. *International Journal of Advanced Academic Research*, 5 (11); 220–41.
- Awuchi CG, Echeta KC (2019). Current Developments in Sugar Alcohols: Chemistry, Nutrition, & Health Concerns of Sorbitol, Xylitol, Glycerol, Arabitol, Inositol, Maltitol, & Lactitol. *Int.J.* Adv. Acad. Res. 5 (11); 1 – 33.
- Awuchi, Chinaza Godswill (2017). Sugar Alcohols: Chemistry, Production, Health Concerns & Nutritional Importance of Mannitol, Sorbitol, Xylitol, & Erythritol. *International Journal of Advanced Academic Research*, 3 (2); 31 – 66.
- Booth, JK; Page, JE; and Bohlmann, J (2017). Hamberger, Björn (edition). "Terpene synthases from Cannabis sativa". *PLOS ONE*. 12 (3): e0173911. doi:10.1371/journal.pone.0173911.
- Boutanaev, MA; Moses, T.; Zi, J.; Nelson, RD; Mugford, S. T.; Peters, JR; and Osbourn, A. (2015). "Investigation of the terpene diversification across multiple sequenced plant genomes". The Proceedings of National Academy of Sciences. 112 (1): E81–E88. doi:10.1073/pnas.1419547112.
- Breitmaier, Eberhard (2006). Terpenes: Flavors, Fragrances, Pharmaca, Pheromones. John Wiley & Sons. pp. 1–13. ISBN 978-3527317868.

- Bringmann, G; Gulder, T; et al. (2011). "Total Synthesis of the Axially Chiral Biaryl Natural Products". *Chemical Reviews*. 111 (2): 563–639. doi:10.1021/cr100155e.
- Brito-Arias, Marco (2007). Synthesis and Characterization of Glycosides. Springer. ISBN 978-0-387-26251-2.
- Castells, Eva and Berenbaum, May R. (2006). "Laboratory Rearing of Agonopterix alstroemeriana, the Defoliating Poison Hemlock (Conium maculatum L.) Moth, and Effects of Piperidine Alkaloids on Preference and Performance". *Environmental Entom ology*. 35 (3): 607–615. doi:10.1603/0046-225x-35.3.607.
- Chiranjibi, P., Sudhakar, R., Dhal, N.K., and Rashmita, D. (2006). Some phytotherapeutic claims by tribals of Rayagada district, Orissa, India. *Ethnobotanical Leaflets*. 10, 189–197.
- Conner W.E. (2009). *Tiger Moths & Woolly Bears—behaviour, ecology, & evolution of the Arctitidae*. New York: Oxford University Press. pp. 1–10. ISBN 0195327373.
- d'Archivio, M; C. Filesi; Varì, R; Scazzocchio, B; and Masella, R (2010). "Bioavailability of the Polyphenols: Status and Controversies". *Int. J. Molecular Sci.*. 11 (4): 1321–1342. doi:10. 3390/ijms11041321.
- Davis, E. M. and Croteau, R (2000). Cyclization enzymes in biosynthesis of monoterpenes, sesquiterpenes, & diterpe nes. *Topics in Current Chem.* 209. pp. 53–95. doi:10.1007/3-540-48146-X_2.
- Eberhard Breitmaier (2006). Terpenes: Flavors, Fragrances, Pharmaca, Pheromones. Wiley-VCH. doi:10.1002/978352760 9949. ISBN 9783527609949.
- EFSA (2011). Scientific Opinion on substantiation of the health claims related to: flavonoids & ascorbic acid in fruit juices, including ... flavonoids (ID". *EFSA Journal*. 9 (4): 2082. doi:10.2903/j.efsa.2011.2082.
- EFSA (2014). "Scientific Opinion on modification of authorisation of a health claim related to cocoa flavanols & maintenance of normal endothelium dependent vasodilation ... Regulation (EC) No 1924/2006". EFSA Journal. 12 (5). May 2014. doi:10.2903/j.efsa.2014.3654.
- Faller, B. A.; and Pandi, T. N. (2011). "Safety & efficacy of vinorelbine in treatment of non-small cell lung cancer". *Clinical Med Insights: Oncology*. 5: 131–44. doi:10.4137/CMO.S5074.
- Feldman, KS and Ensel, SM (1994). "The ellagitannin chemistry. Preparative & mechanistic studies of the biomimetic oxidative coupling of galloyl esters". *J. Am. Chem. Soc.* 116 (8): 3357–3366. doi:10.1021/ja00087a022.
- Feldman, KS; Lawlor, DM; and Sahasrabudhe, K (2000). "The ellagitannin chemistry. The evolution of a three-component coupling strategy for synthesis of dimeric ellagitannin coriariin A & a dimeric gallotannin analogue". *Journal of Organic Chemistry*. 65 (23): 8011–19. doi:10.1021/j00010936.
- Feldman, SK (2004). "Recent progress in the ellagitannin chemistry". *Phytochemistry*. 66 (17): 1984 to 2000. doi:10.1016 /j.phytochem.2004.11.15.
- Fox, Eduardo G. P.; Wu, Xiaoqing; Wang, Lei; Chen, Li; Lu, Yong-Yue; and Xu, Yijuan (2019). Queen venom isosolenopsin A delivers rapid incapacitation of fire ant competitors. *Toxicon.* 158: 77–83. doi:10.1016/j.toxicon.2018.11.428. IS SN 0041-0101.
- Fox, Eduardo G. P.; Xu, Meng; Wang, Lei; Chen, Li; and Lu, Yong-Yue (2018). "Gas-chromatography and UV-spectroscopy of Hymenoptera venoms obtained by trivial centrifugation". *Data in Brief.* 18: 992–998. doi:10.1016/j.dib.2018.03.101. ISSN 2352-3409.

- Fox, Eduardo Gonçalves Paterson (2016). Gopalakrishnakone, P.; Calvete, Juan J. (eds.), "Venom Toxins of Fire Ants", Venom Genomics and Proteomics: Venom Genomics and Proteomics, Springer Netherlands, pp. 1–16, doi:10.1007/978-94-007-6649-5_38-1 (inactive 2019-08-30), ISBN 9789400766495
- Fox, Eduardo PG; Xu, M; Wang, L; Chen, L; and Lu, YY (2018). "Speedy milking of fresh venom from aculeate hymenopterans". *Toxicon*. 146: 120–123. doi:10.1016/j.toxicon. 2018.02.050. ISSN 0041-0101.
- Gansäuer, A; Justicia, J; Fan, CA; Worgull, D; and Piestert, F (2007). "Reductive C—C bond formation after the epoxide opening via the electron transfer". In Krische, Michael J. (edition). Metal Catalyzed Reductive C—C Bond Formation: ... Topics in Current Chemistry. 279. Springer Science and Business Media. 25–52. doi:10.1007/128_2007_130.
- Gleadow, RM and Møller, BL (2014). "Cyanogenic glycosides: synthesis, physiology, and phenotypic plasticity". *Annual Review of Plant Biology*. 65: 155–85. doi:10.1146/annurev-arplant-050213-040027.
- Gonçalves Paterson Fox, Eduardo; Russ Solis, Daniel; Delazari dos Santos, Lucilene; Aparecido dos Santos Pinto, Jose Roberto; Ribeiro da Silva Menegasso, Anally; Cardoso Maciel Costa Silva,
- Gross, Paul (2009). "New Roles for the Polyphenols. A 3-Part Report on the Current Regulations & the State of Science". Nutraceuticals World.
- György M, Miklós N, and Viktor A (2002). *Pesticide chemistry*, Elsevier, 2002, page 21–22 ISBN 0-444-98903-X.
- Halliwell, B (2007). "The dietary polyphenols: Good, bad, or indifferent for your health?". *Cardiovascular Research.* 73 (2): 341–47. doi:10.1016/j.cardiores.2006.10.04.
- Haslam, E.; and Cai, Y. (1994).Plant polyphenols (vegetable tannins): Gallic acid metabolism. *Natural Product Reports*. 11 (1): 41–66. doi:10.1039/NP9941100041.
- Haslam, E; Lilley, HT; Cai, Y; Martin, R; and Magnolato, D (1989). "Traditional herbal medicines-- role of the polyphenols". *Planta Med.* 55 (1): 1–8. doi:10.1055/s-2006-961764.
- Hättenschwiler, S and Vitousek, P M (2000). "The role of polyphenols in the terrestrial ecosystem nutrient cycling". *Trends in Ecology and Evolution*. 15 (6): 238–243. doi:10.1016/S0169-5347(00) 01861-9
- Hertweck, C. (2009). "The Biosynthetic Logic of Polyketide Diversity". *Angewandte Chemie International Edition*. 48 (26): 4688–716.doi:10.1002/anie.200806121.
- Hirata, K.; Miyamoto, K.; and Miura, Y. (1994). "Catharanthus roseus L.: Production of Vindoline & Catharanthine in Multiple Shoot Cultures". In ... Biotechnology in Agriculture & Forestry 26. Medicinal & Aromatic Plants. VI. Springer-Verlag. pp. 46– 55. ISBN 9783540563914.
- Hufnagel JC, and Hofmann T (2008). "The orosensory-directed identification of astringent mouthfeel & bitter-tasting compounds in red wine". *J Agric Food Chem.* 56 (4): 1376–1386. doi:10.1021/jf073031n.
- Itoh, T; Chika, J; et al. (1996). "Synthesis of trideca-O-methyl-alphapedunculagin. The diastereo-favoritism studies on intramolecular ester-cyclization of axially chiral diphenic acids with carbohydrate core". *Journal Organic Chemistry*. 61 (11): 3700–3705. doi:10.1021/jo950969j.
- IUPAC (2009). "Glycosides". IUPAC Gold Book Glycosides. 2009. doi:10.1351/goldbook.G02661. ISBN 978-0-9678550-9-7.
- Jindal K. K. and Sharma R. C. (2004). Recent trends in horticulture in the Himalayas. Indus Publishing. ISBN 978-81-7387-162-7.
- Khanbabaee, K and Grosser, M (2003). "Efficient total synthesis of pedunculagin by using twofold intramolecular double

- esterification strategy". European Journal of Organic Chemistry. 2003 (11): 2128–2131. doi:10.1002/ejoc.200300006.
- Kittakoop P, Mahidol C, Ruchirawat S (2014). "Alkaloids as important scaffolds in therapeutic drugs for the treatments of cancer, tuberculosis, and smoking cessation". *Curr Top Med Chem.* 14 (2): 239–252. doi:10.2174/156802661366613121610504 9.
- Krasnow, MN and Murphy, MT (2004). "Polyphenol Glucosylating Activity in the Cell Suspensions of Grape (*Vitis vinifera*)". *J Agric and Food Chem.* 52 (11): 3467–72. doi:10.1021/jf035234r.
- Krohn, K.; I. Ahmed; John, M.; Letzel, CM; and Kuck, D. (2010).
 "Stereoselective Synthesis of the Benzylated Prodelphinidins & their Diastereomers with the use of the Mitsunobu Reaction in Preparation of their Gallocatechin Precursors". European Journal of Organic Chemistry. 2010 (13): 2544–2554. doi:10.1002/ejoc.2 01000053.
- Kumari, I.; Ahmed, M.; and Akhter, Y. (2017). "The evolution of catalytic microenvironment governs the substrate & product diversity in trichodiene synthase & other terpene fold enzymes". *Biochimie*. 144: 9–20. doi:10.1016/j.biochi.2017.10.03.
- Lamorde, M., Tabuti, J.R.S., Obua, C., Kukunda-Byobona, C., Lanyero, H., Byakika- Kibwika, P., Bbosa, S.G., Lubega, A., Ogwal-Okeng, J., Ryan, M.,Waako, P.J., and Merry, C. (2010). Medicinal plants used by the traditional medicine practitioners in treatment of HIV/AIDS & related conditions in Uganda. *Journal* of Ethnopharmacology 130, 43–53.
- Lattanzio V. *et al.* (2006). Role of phenolics in resistance mechanisms of plants against the fungal pathogens & insects (and references therein). *Phytochemistry*: Advances in Research, 23-67. ISBN 81-308-0034-9.
- Leland J. C. (2006). Natural Products from Plants2nd Edition. CRC, 2006, p. 30 ISBN 0-8493-2976-0
- Lesschaeve I, and Noble CA (2005). "Polyphenols: the factors influencing their sensory properties & their effects on food & beverage preferences". *American Journal of Clinical Nutrition*. 81(1): 330S–35S. doi:10.1093/ajcn/81.1.330S.
- Lindhorst, T.K. (2007). Essentials of Carbohydrate Chemistry and Biochemistry. Wiley-VCH. ISBN 978-3-527-31528-4.
- Martin, D. M.; Gershenzon, J.; and Bohlmann, J. (2003). "The Induction of Volatile Terpene Biosynthesis & Diurnal Emission by Methyl Jasmonate in Foliage of Norway Spruce". *Plant Physiology*. 132 (3): 1586–99. doi:10.1104/pp.103.021196.
- Mello, L; Sotomayor, Maria Del P T; Kubota, L T (2003). "HRP-based amperometric biosensor for polyphenols determination in vegetables extract". *Sensors & Actuators B: Chemical*. 96 (3): 636–645. doi:10.1016/j.snb.2003.07.08.
- Mennen L.; et al. (2005). "Risks and Safety of Polyphenol Consumption". *American Journal of Clinical Nutrition*. 81 (1): 3265–3295. doi:10.1093/ajcn/81.1.326S.
- Milazzo, S and Horneber, M (2015). "Laetrile treatment for cancer".

 Cochrane Database of Systematic Reviews (4):

 CD005476. doi:10.1002/14651858.CD05476.pub4.
- Morcillo, S P.; Miguel, D; Campaña, Araceli G.; Cienfuegos, L Álvarez de; Justicia, J; and Cuerva, J M. (2014). "Recent applications of the Cp_2TiCl in the natural product synthesis". Organic Chemistry Frontiers. 1 (1): 15–33. doi:10.1039/c3qo00024a.
- Nelson, TD and Meyers, IA (1994). "A Rapid Total Synthesis of Ellagitannin [sic]". *J. Org. Chem.* 59 (9): 2577–80. doi:10.1021/jo00088a046.
- Ngo, Q A; Roussi, F; Cormier, A; Thoret, S; Knossow, M; Guénard, D; Guéritte, F (2009). "Synthesis & biological evaluation of Vinca alkaloids & phomopsin hybrids". *J Medicinal Chem.* 52 (1): 134–

- 42. doi:10.1021/jm801064y.
- Osman, A. M.; Wong, K. K. Y.; and Fernyhough, A. (2006). "The ABTS radical-driven oxidation of the polyphenols: Isolation and structural elucidation of covalent adducts". *Biochemical & Biophysical Research Communications*. 346 (1): 321–29. doi:10. 1016/j.bbrc.2006.5.118.
- Owen, WR.; Haubner, R.; Hull, EW; Erben, G.; Spiegelhalder, B.; Bartsch, H.; and Haber, B. (2003). "Isolation & structure elucidation of major individual polyphenols in carob fibre". *Food & Chemical Toxicology*. 41 (12): 1727–1738. doi:10.1016/S0278-6915(03)00200-X.
- Pan, X (2003). "The microwave-assisted extraction of tea polyphenols & tea caffeine from green tea leaves". *Chemical Engineering & Processing*, 42 (2): 129–33. doi:10.1016/S0255-2701(02)00037-5.
- Papadopoulou, Athina and Frazier, Richard A (2004). "Characterization of protein–polyphenol interactions". *Trends in Food Sci and Technol.* 15 (3–4): 186–190. doi:10.1016/j.tifs.2003.09.17.
- Patra, AK and Saxena, J (2009). "The effect & mode of action of saponins on the microbial populations & fermentation in the rumen & ruminant production". Nutr Res Rev. 22 (2): 204– 209. doi:10.1017/S0954422409990163.
- Pazouki, L. and Niinemets, Ü. (2016). "The Multi-Substrate Terpene Synthases: Their Occurrence & Physiological Significance". Frontiers in Plant Science. 7: 1019. doi:10.3389/fpls.2016.01019.
- Pianaro, Adriana; Fox, Eduardo G. P.; Bueno, Odair C.; Marsaioli, Anita J. (2012). "Rapid configuration analysis of the solenopsins". *Tetrahedron: Asymmetry*. 23 (9): 635–642. doi:10. 1016/j.tetasy.2012.05.005. ISSN 0957-4166.
- Pichersky, E. (2006). "Biosynthesis of the Plant Volatiles: Nature's Diversity & Ingenuity". *Science*. 311 (5762): 808–811.doi:10.1126/science.1118510.
- Popa, V; Dumitru, M; Volf, I; and Anghel, N (2008). "Lignin & polyphenols as allelochemicals". *Industrial Crops & Products*. 27 (2): 144–149. doi:10.1016/j.indcrop.2007.07.19.
- Pouysegu, L.; Deffieux, D.; et al. (2011). "Synthesis of ellagitannin natural products". *Nat. Prod. Rep.* 28 (5): 853–874. doi:10.1039/c0np00058b.
- Pulido, R.; Bravo, L.; Saura-C, F. (2000). "The Antioxidant Activity of the Dietary Polyphenols As Determined by a Modified Ferric Reducing/Antioxidant Power Assay". *Journal of Agricultural and Food Chemistry*. 48 (8): 3396–3402. doi:10.1021/jf9913458. hdl :10261/112476.
- Quideau S (2011). "Why bother with polyphenols?". Groupe Polyphenols. Retrieved 26 March 2014.
- Quideau, PS.; Deffieux, D.; Douat-Casassus, LC; and Pouységu, L. (2011).Plant Polyphenols: the Chemical Properties, Biological Activities, & Synthesis. Angewandte Chemie Inter'l Edition. 50 (3): 586–621. doi:10.1002/anie.201000044.
- Quideau, S. (2011b). "Organic chemistry: Triumph for unnatural synthesis". *Nature*. 474 (7352): 459–460. doi:10.1038/474459a.
- Rafael; Sergio Palma, Mario; Correa Bueno, Odair; de Alcântara Machado, Ednildo (2013). "A simple, rapid method for extraction of whole fire ant venom (Insecta: *Formicidae: Solenopsis*)". *Toxicon.* 65: 5–8. doi:10.1016/j.toxicon.2012.12.009.
- Raj, K B (2004). A Text Book of Organic Chemistry. 4th Edition, New Age International, 2004, p. 644 ISBN 81-224-1459-1
- Robbers JE, Speedie KM, and Tyler VE (1996). "Chapter 9: Alkaloids". Pharmacognosy and Pharmacobiotechnology. Philadelp hia: Lippincott, Williams & Wilkins. pp. 143–185. ISBN 978-0683085006.

- Robert Alan Lewis (1998). Lewis' dictionary of toxicology. CRC Press, 1998, p. 51 ISBN 1-56670-223-2
- Roberts, Margaret F. (1998). Alkaloids: Biochemistry, Ecology, and Medicinal Applications. Wink, Michael. Boston, MA: Springer US. ISBN 9781475729054. OCLC 851770197.
- Russo P, Frustaci A, Del Bufalo A, Fini M, Cesario A (2013). "Multitarget drugs of plants origin acting on Alzheimer's disease". Curr Med Chem. 20 (13): 1686–93. doi:10.2174/0929 867311320130008.
- Sarkar S, Zaidi S, Chaturvedi AK, Srivastava R, Dwivedi PK, and Shukla R. (2015). Search for a herbal medicine: Antiasthmatic activity of methanolic extract of *Curcuma longa. Journal of Pharmacognosy and Phytochemistry*. 2015; 3: 59-72.
- Shrestha PM and Dhillion S. S. (2003). Medicinal plant diversity & use in highlands of Dolakha district, Nepal. *J Ethnopharmacol*. 2003; 86(1): 81-96.
- Snyder, S.A.; Gollner, A.; and Chiriac, M.I. (2011). "Regioselective reactions for programmable resveratrol oligomer synthesis". *Nature*. 474 (7352): 461–466. doi:10.1038/nature101 97.
- Spiller G. A. (1997). *Caffeine*, CRC Press, 1997 ISBN 0-8493-2647-8
 Steenackers, B.; De Cooman, L.; and De Vos, D. (2015). "The chemical transformations of the characteristic hop secondary metabolites in relation to the beer properties & the brewing process: A review". *Food Chem.* 172: 742–56. doi:10.1016/j.foodchem.2014.9.139.
- Sun, Hong-Xiang; Xie, Yong; and Ye, Yi-Ping (2009). "Advances in the saponin-based adjuvants". *Vaccine*. 27 (12): 1787–96. doi:10.1016/j.vaccine.2009.01.91.
- Tabuti, J.R.S., Dhillion, S.S., and Lye, K.A., (2003). Traditional medicine in Bulamogi County, Uganda. Its practitioners, users & viability. J. Ethnopharmacol. 85, 119–129.
- Tabuti, J.R.S., Kukunda, C.B., and Waako, W.J. (2010). Medicinal plants used by the traditional medicine practitioners in treatment of tuberculosis & related ailments in Uganda. *J. Ethnopharmacol*. 127, 130–136.
- Tanner, G J; Francki, K T; Abrahams, S; Watson, J M; Larkin, P J; and Ashton, A R (2003)."Proanthocyanidin Biosynthesis in Plants". *J. Biolog. Chem.* 278 (34): 31647–31656. doi:10.107 4/jbc.M302783200.
- Thimmappa, R.; Geisler, K.; T. Louveau; O'Maille, P.; and Osbourn, A. (2014). "The triterpene biosynthesis in plants". *Annual Rev Plant Biol.* 65: 225–57. doi:10.1146/annurev-arplant-050312-120229.
- Thomas Acamovic; Colin S. Stewart; and T. W. Pennycott (2004). *Poisonous plants and related toxins, Vol. 2001. CABI.* p. 362. ISBN 978-0-85199-614-1.
- Touchard, Axel; Aili, Samira; Fox, Eduardo; Escoubas, Pierre; Orivel, Jérôme; Nicholson, Graham; and Dejean, Alain (2016). "The Biochemical Toxin Arsenal from Ant Venoms". *Toxins*. 8 (1): 30. doi:10.3390/toxins8010030. ISSN 2072-6651.
- US Food and Drug Administration (2017). "Guidance for Industry: Food Labeling; Nutrient ... Compliance Guide". Center for Food Safety & Applied Nutrition, US Food & Drug Administration.
- Walker, Richard B. and Everette, J D (2009). "The Comparative Reaction Rates of Various Antioxidants with the ABTS Radical Cation". *J. Agric Food Chem.* 57 (4): 1156–1161. doi:10.10 21/jf8026765.
- Zelena, K.; Hardebusch, B.; B. Hülsdau; Berger, R. G.; and Zorn, H. (2009). "The Generation of Norisoprenoid Flavors from the Carotenoids by Fungal Peroxidases". *J. Agric Food Chem.* 57 (21): 9951–9955. doi:10.1021/jf901438m.